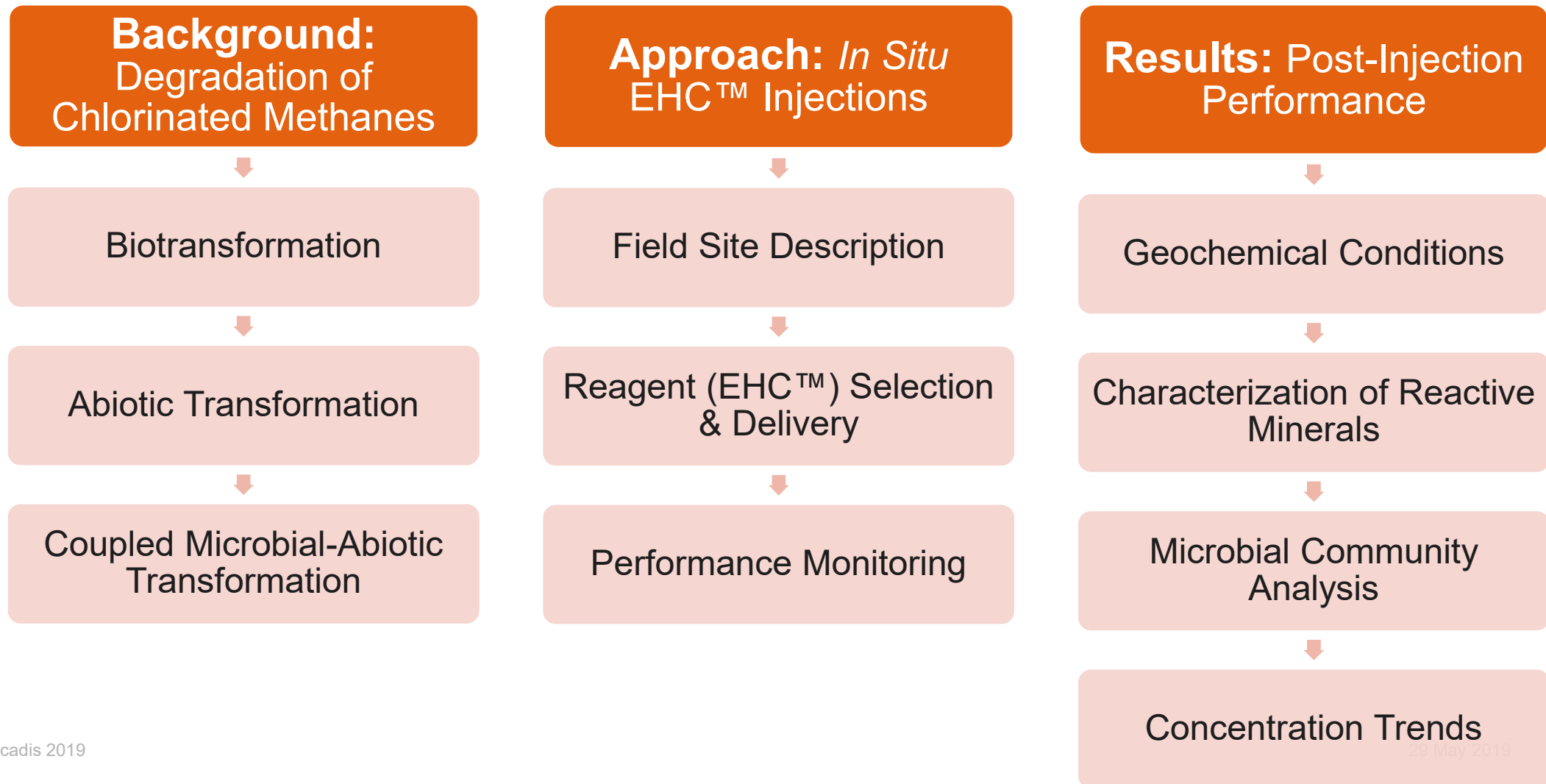


VERIFIED CARBON TETRACHLORIDE AND CHLOROFORM TRANSFORMATION VIA BIOTIC, ABIOTIC, AND REACTIVE IRON SULFIDE MECHANISMS

Shandra D. Justicia-León, Jennifer Martin Tilton, Matthew Schnobrich, Craig Divine,
Shannon M. Ulrich, David Liles, and Dora Taggart

Battelle Fifth International Symposium on Bioremediation and Sustainable Technologies
April 2019 – Baltimore, Maryland

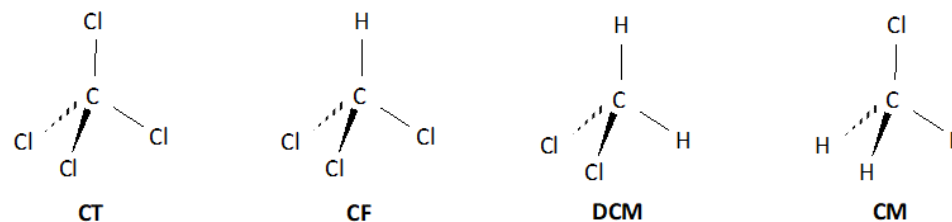
Agenda



Degradation of chlorinated methanes

- Biotransformation
 - ✓ Cometabolic biotransformation
 - ✓ Catabolic biotransformation: sources of carbon/energy or respiratory electron acceptors
- Abiotic transformation
 - ✓ Degradation by hydrolysis (e.g., via contact with zero-valent metals)
 - ✓ CT more amenable to abiotic transformation
- Coupled microbial-abiotic transformation of chlorinated methanes
 - ✓ Abiotic reactions catalyzed by reactive mineral species (e.g., iron sulfur minerals) typically depend on microbial activity, which generates the reactive species/surfaces

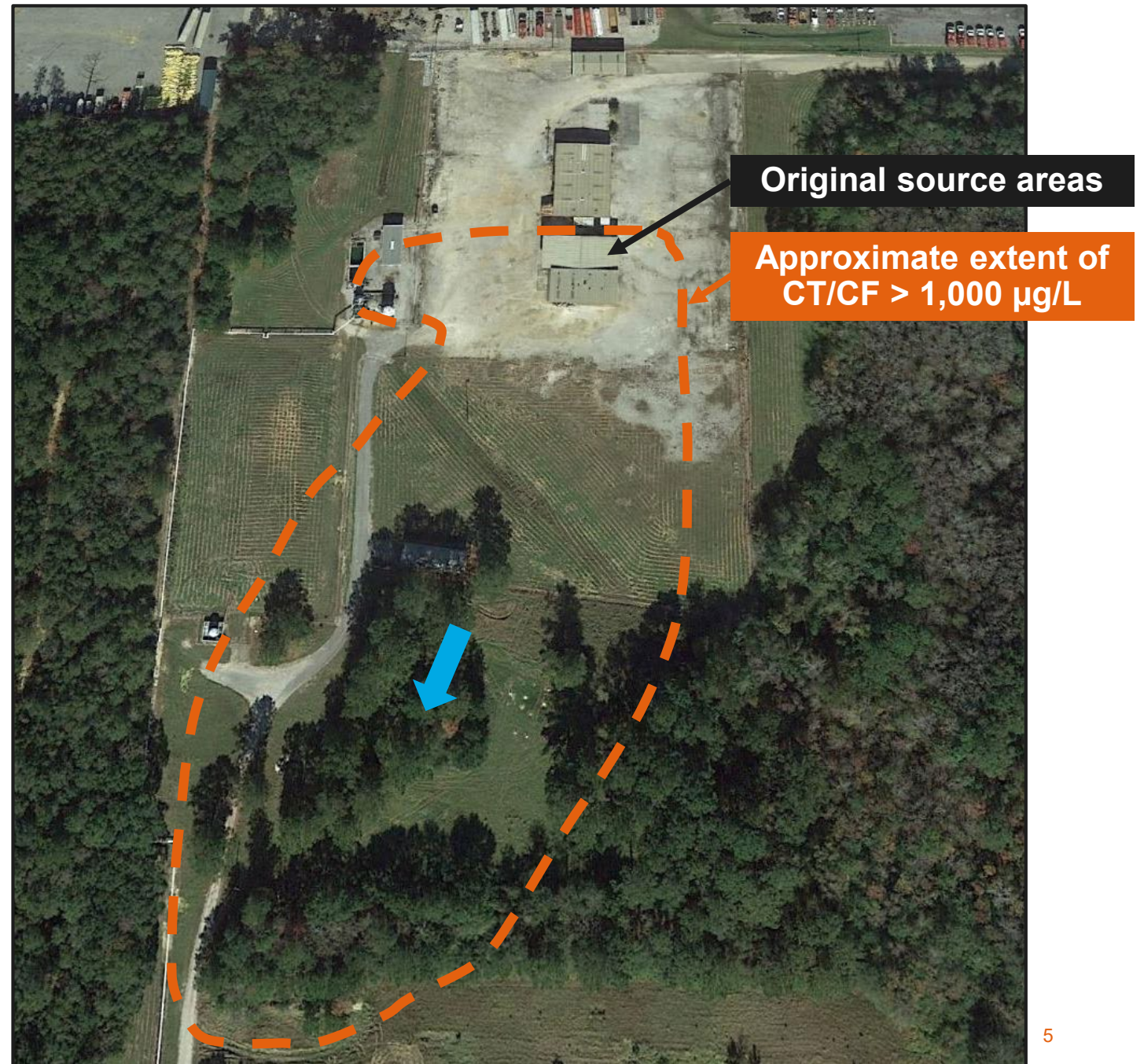
organohalide respiration



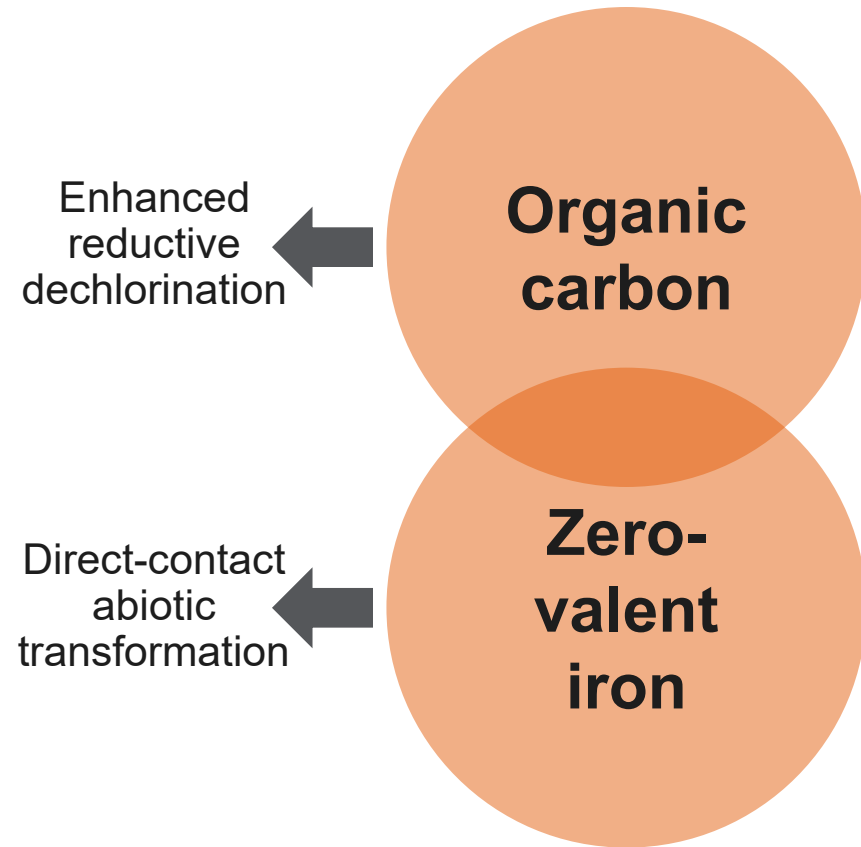
Approach: *In Situ* EHC™ Injections

Field Site Description

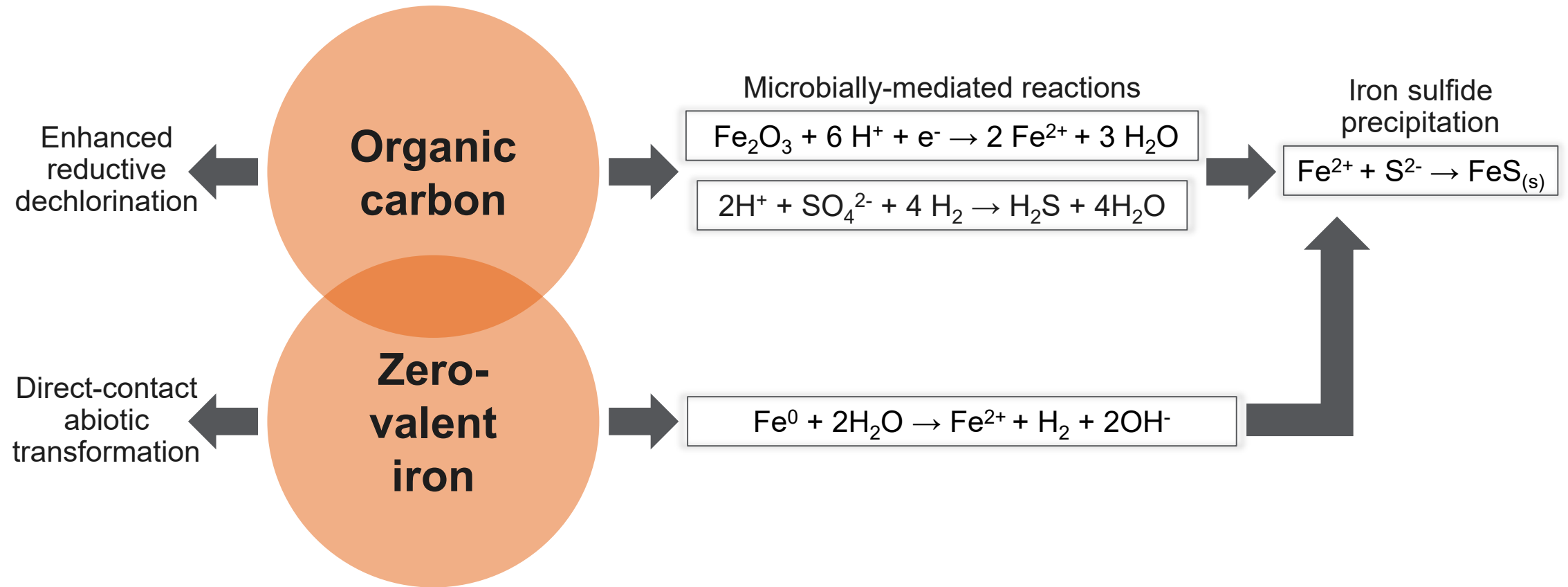
- Primary contaminants of concern: CT and CF
- Max chlorinated methane concentration ~ 20 mg/L
- Co-disposed sulfur-containing compounds
- Naturally high iron
- Remedial approach: *in situ* EHC™ injections



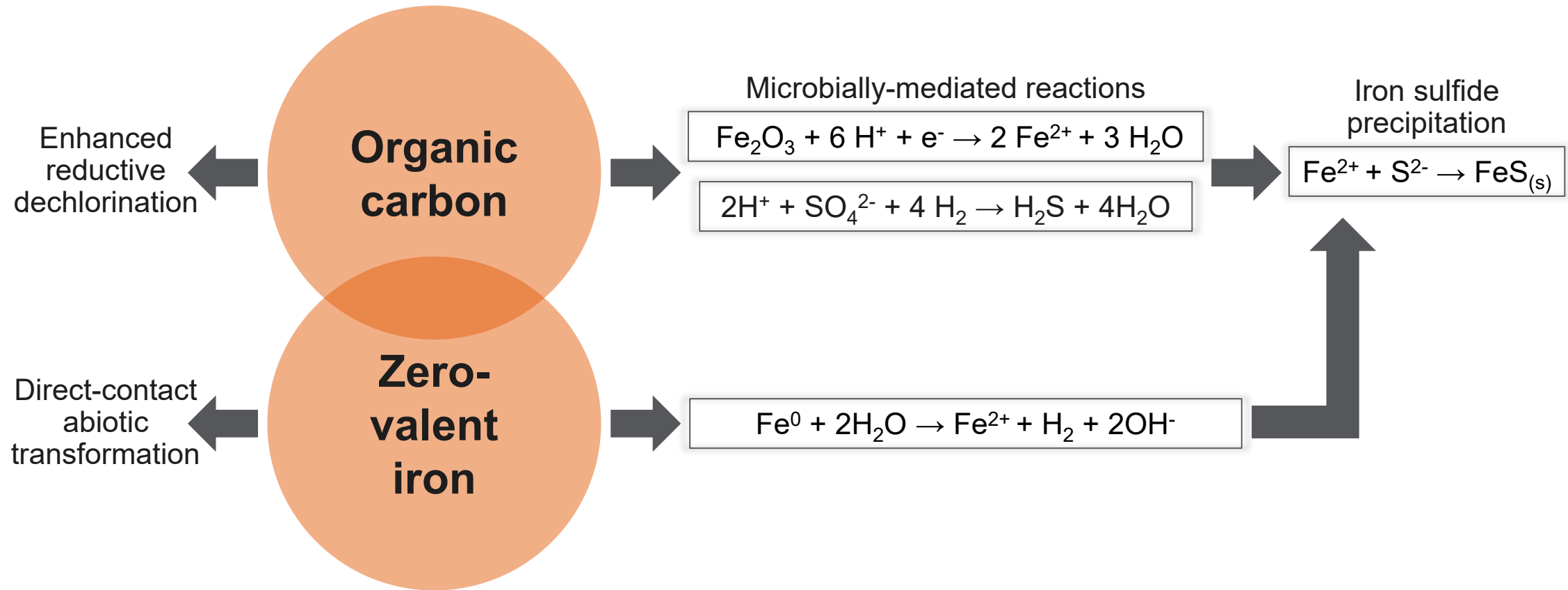
Reagent Selection: EHC™



Reagent Selection: EHC™



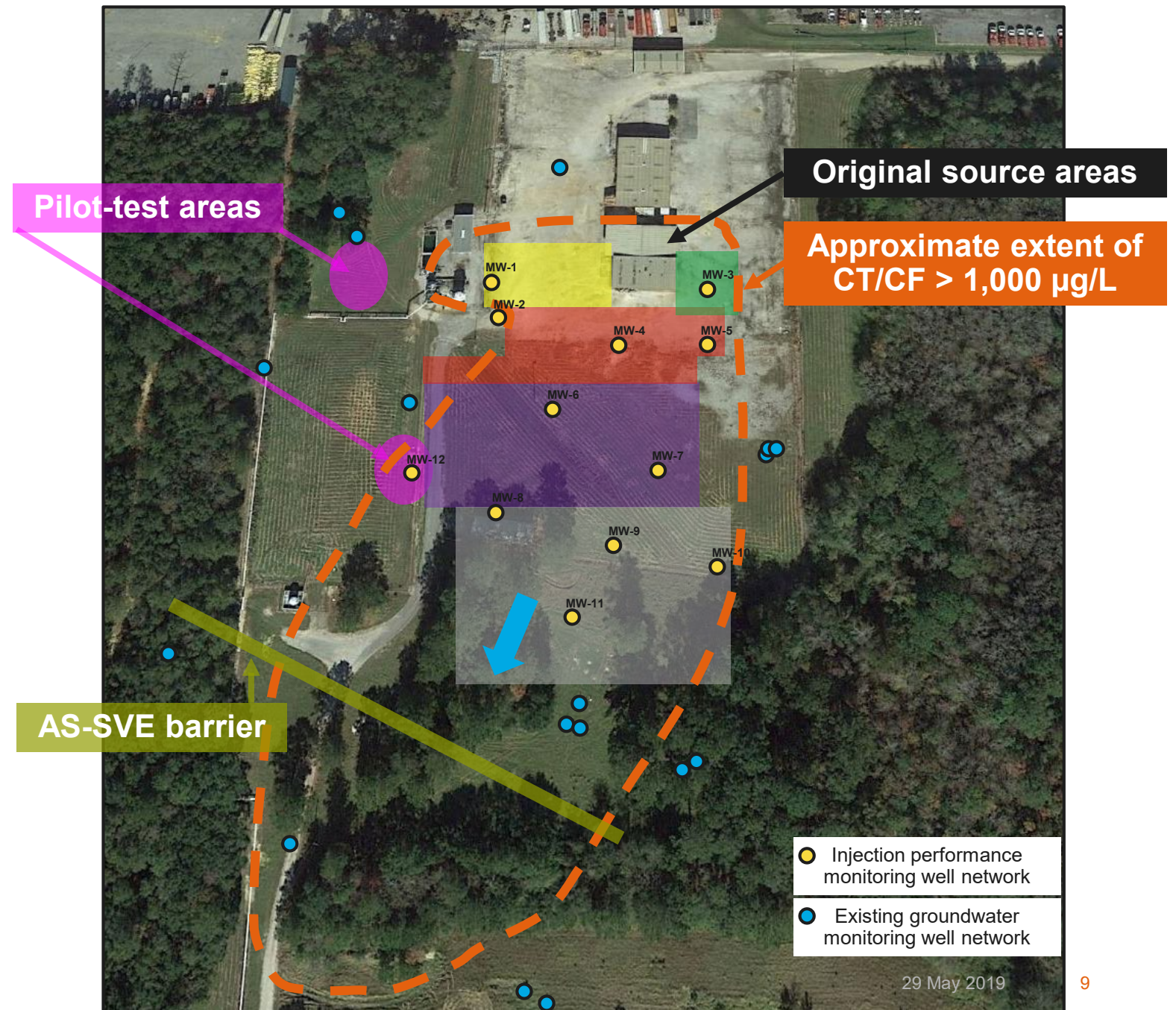
Reagent Selection: EHC™



Expanded injection radius of influence & increased degradation capacity beyond direct utilization of EHC™

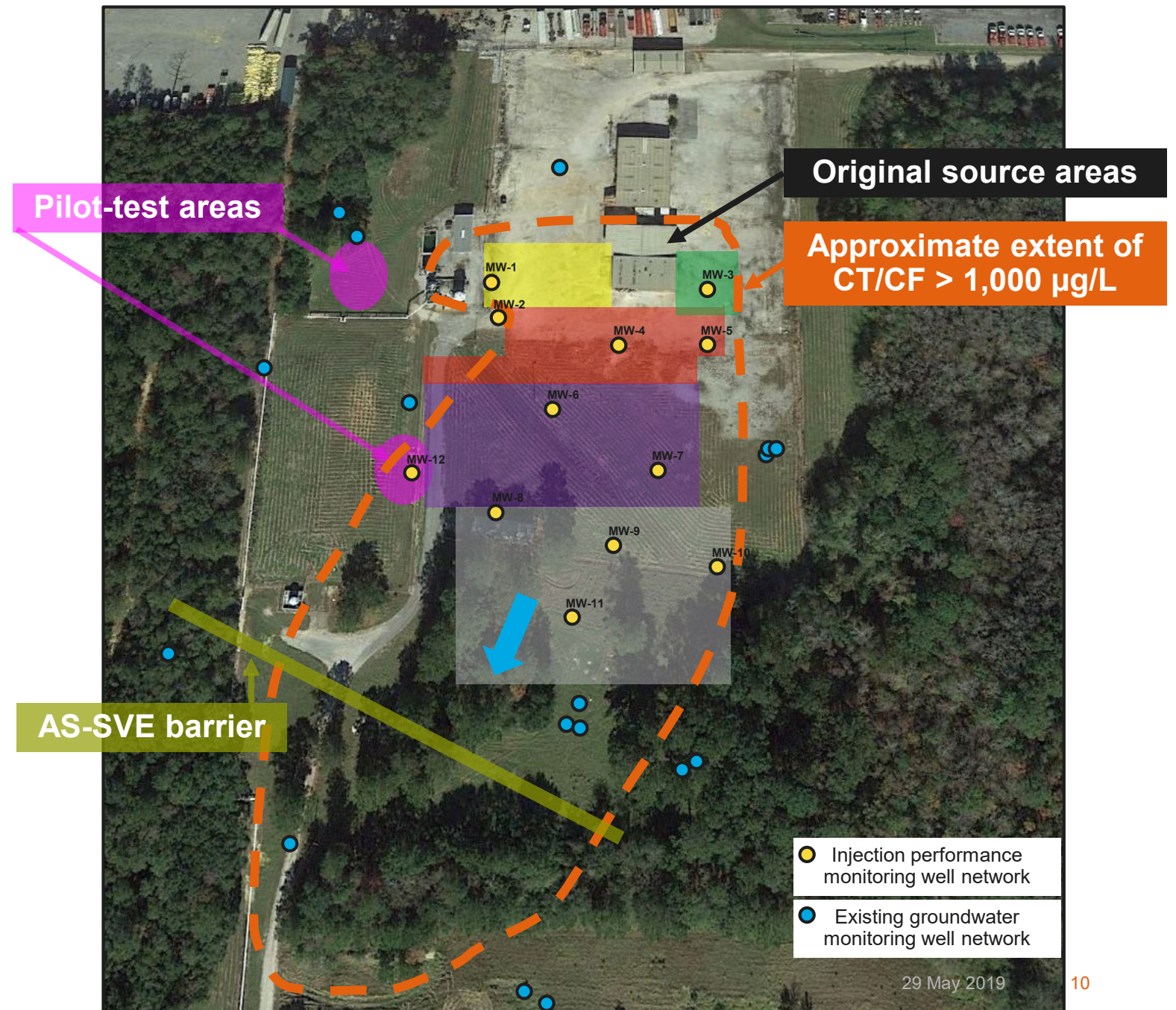
EHC™ Delivery

- Pilot-scale delivery
May 2017
- Full-scale delivery
June-August 2018
- Direct-push injection
of ~ 274,000 lbs
EHC™
- ~ 300 injection points
- Performance
monitoring August-
December 2018



Performance Monitoring

- Assessment of geochemical parameters
- Characterization of iron sulfide minerals
- Characterization of microbial community
- Evaluation of concentration trends for CT, CF and degradation products



Performance Monitoring

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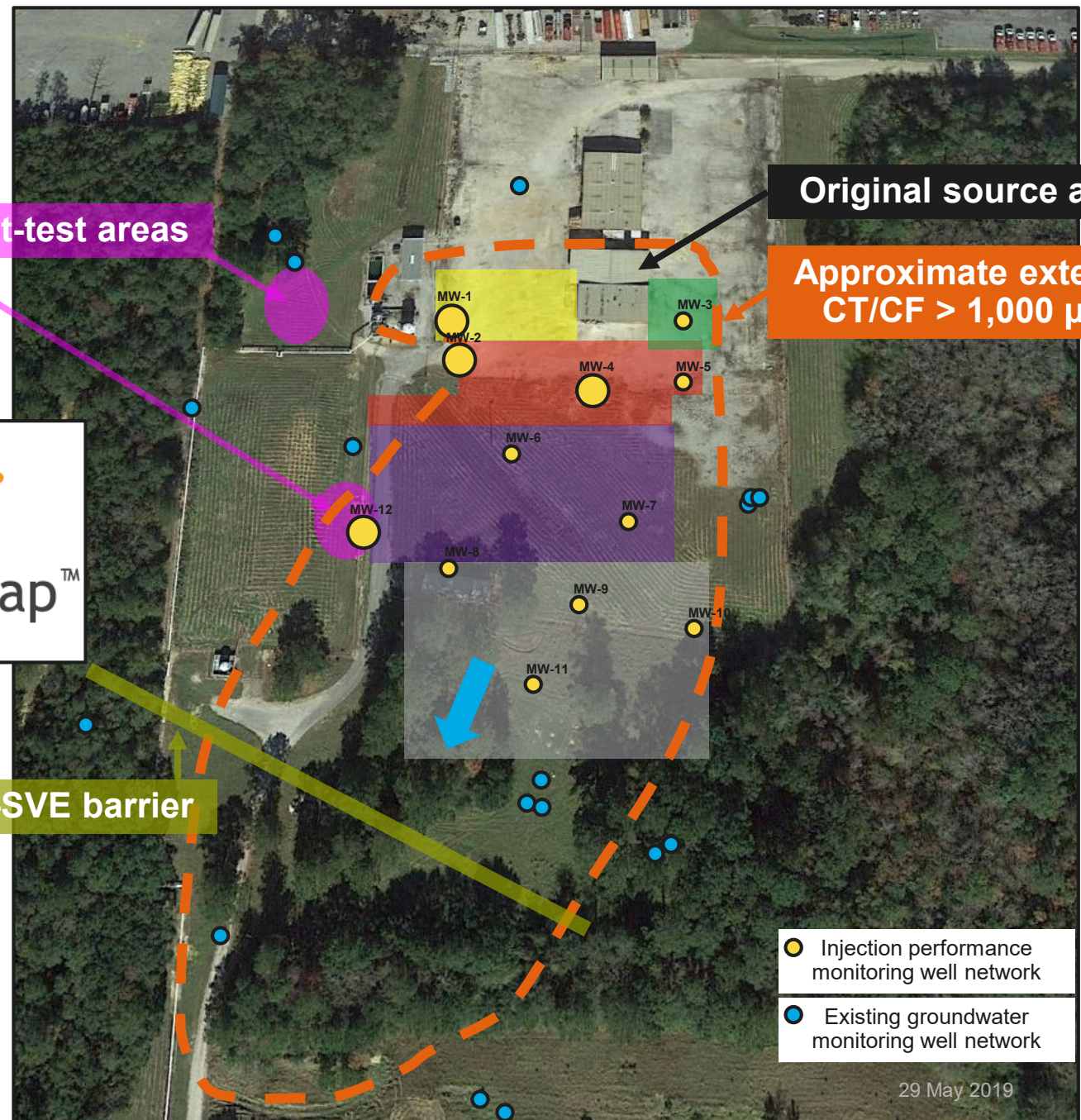
Pilot-test areas

Original source areas

Approximate extent of CT/CF > 1,000 µg/L

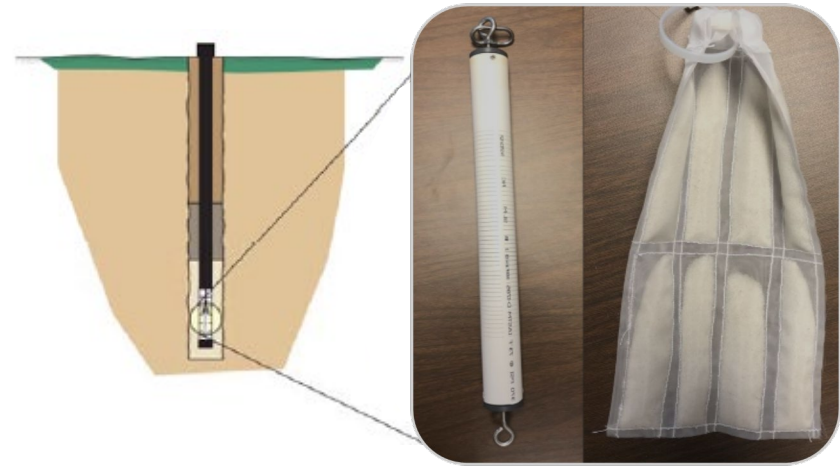
AS-SVE barrier

- Injection performance monitoring well network
- Existing groundwater monitoring well network



New Monitoring Tool: Min-Traps™

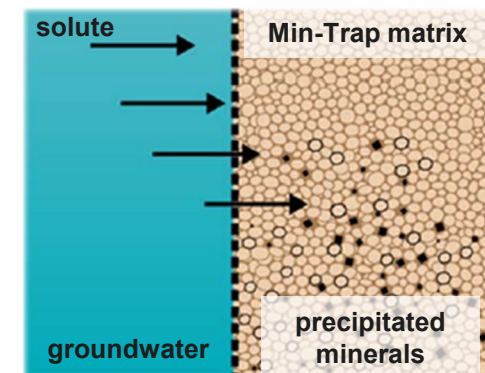
- Collect minerals forming *in situ* using existing monitoring well network
 - ✓ Representative of conditions in higher-flux zones
- Porous medium inside mesh acts as a carrier for target minerals
 - ✓ Medium is customizable
- Inexpensive, easily repeated



Deployed in standard 2"+ monitoring well

Slotted PVC casing & porous medium in permeable mesh

Min-Traps can conclusively document in situ formation of specific minerals



New Monitoring Tool: Min-Traps™

More on Min-Traps!



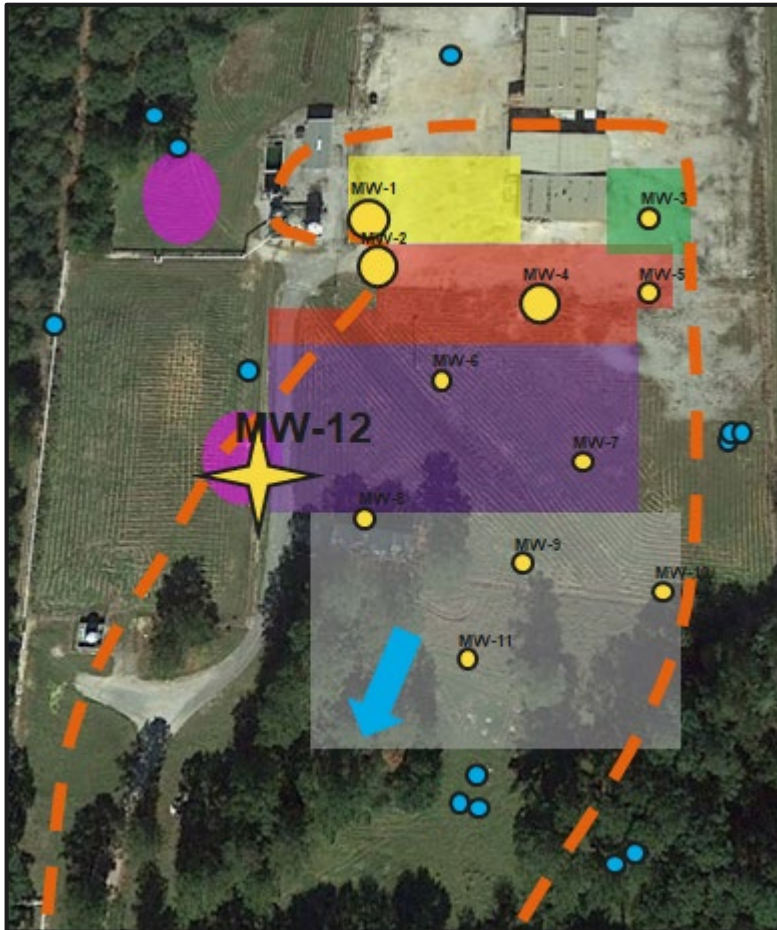
**Wednesday, 4:45 pm, D Session
(Holiday Ballroom 4)**

Documenting *In Situ* Reactive Iron
Mineral Formation without Drilling: A
New Monitoring Well-Based Sampling
Approach

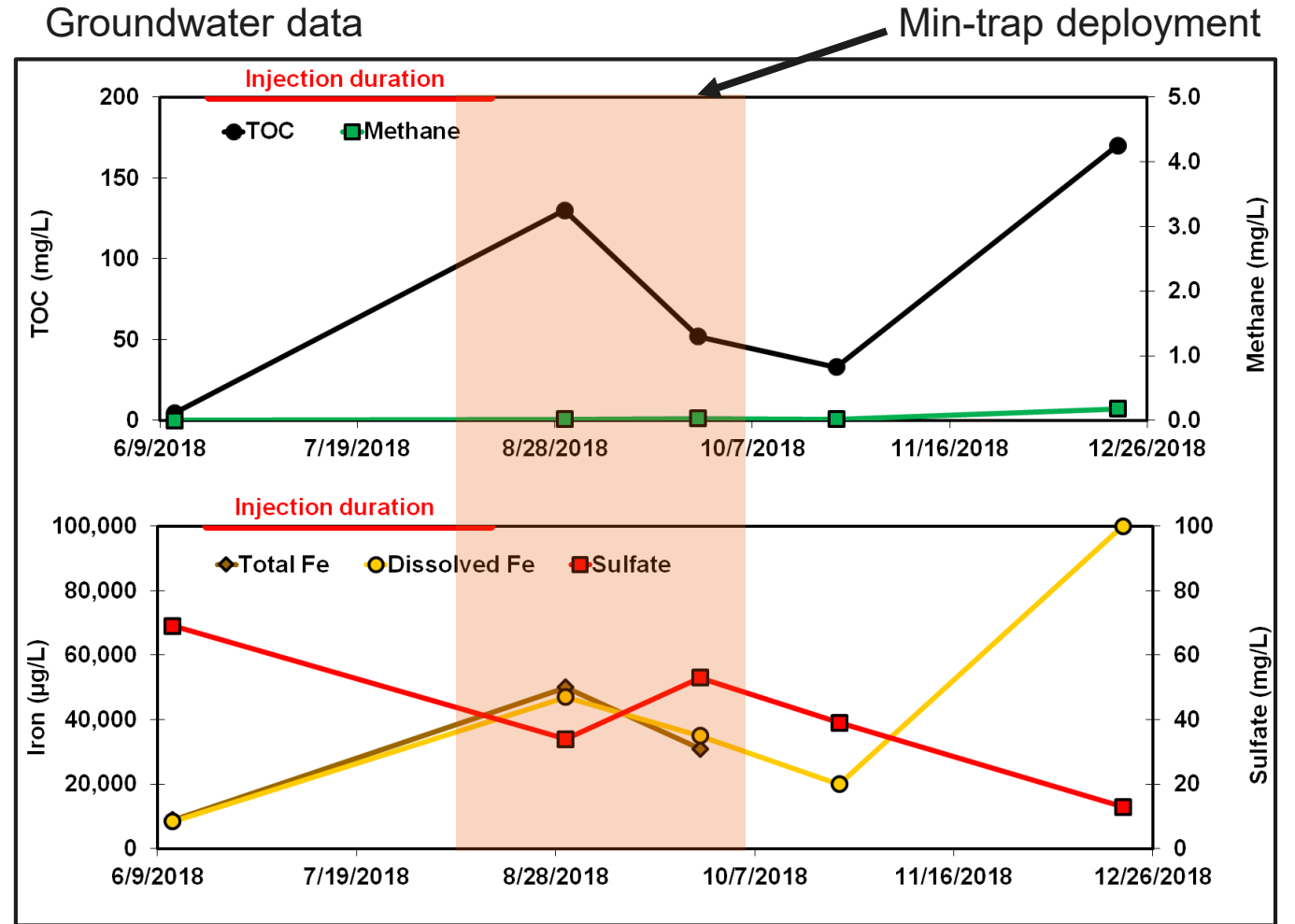
Jennifer Martin Tilton (Arcadis)

Results: Post-Injection Performance

Post-Injection Performance



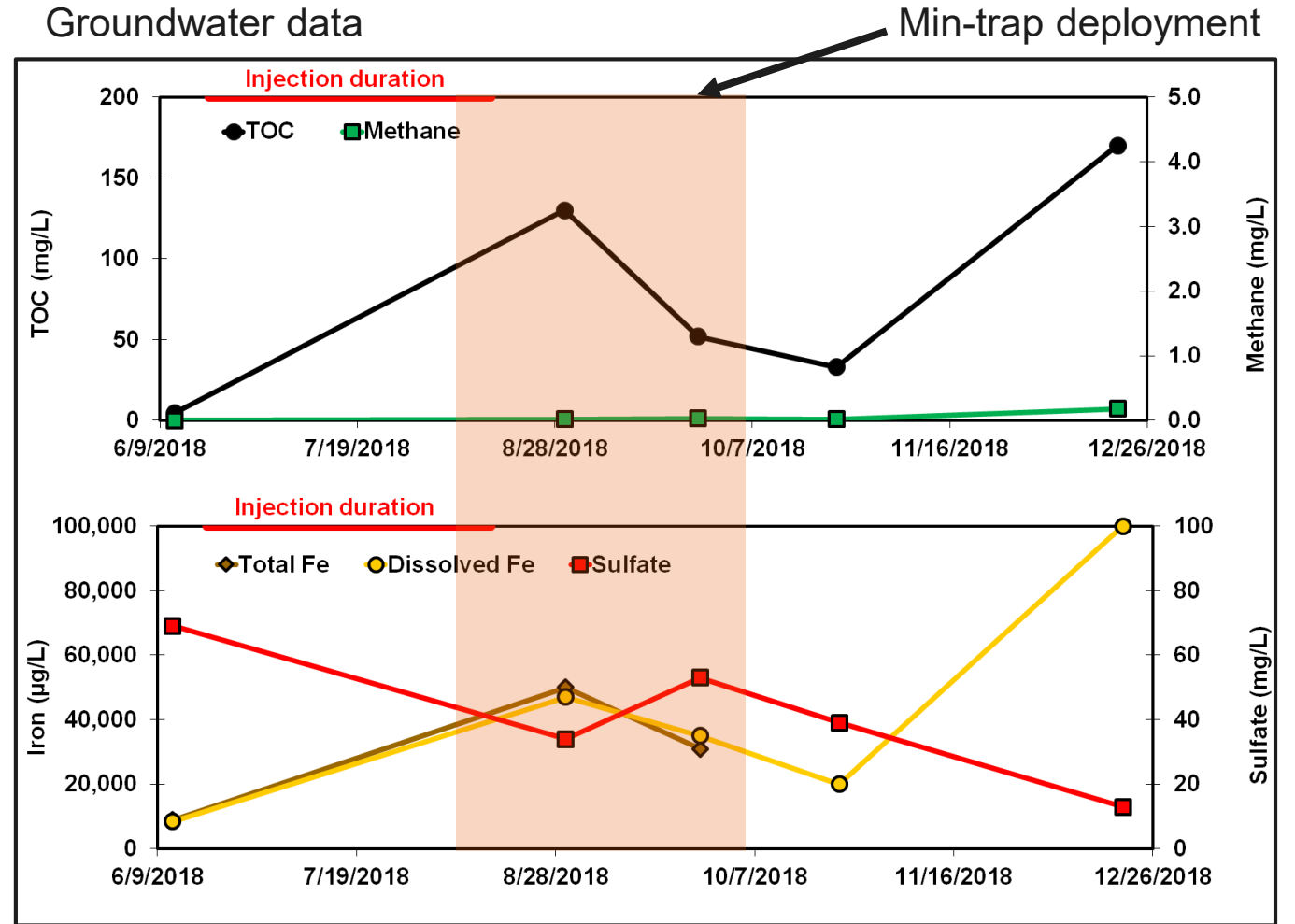
Groundwater data



Post-Injection Performance



Groundwater data

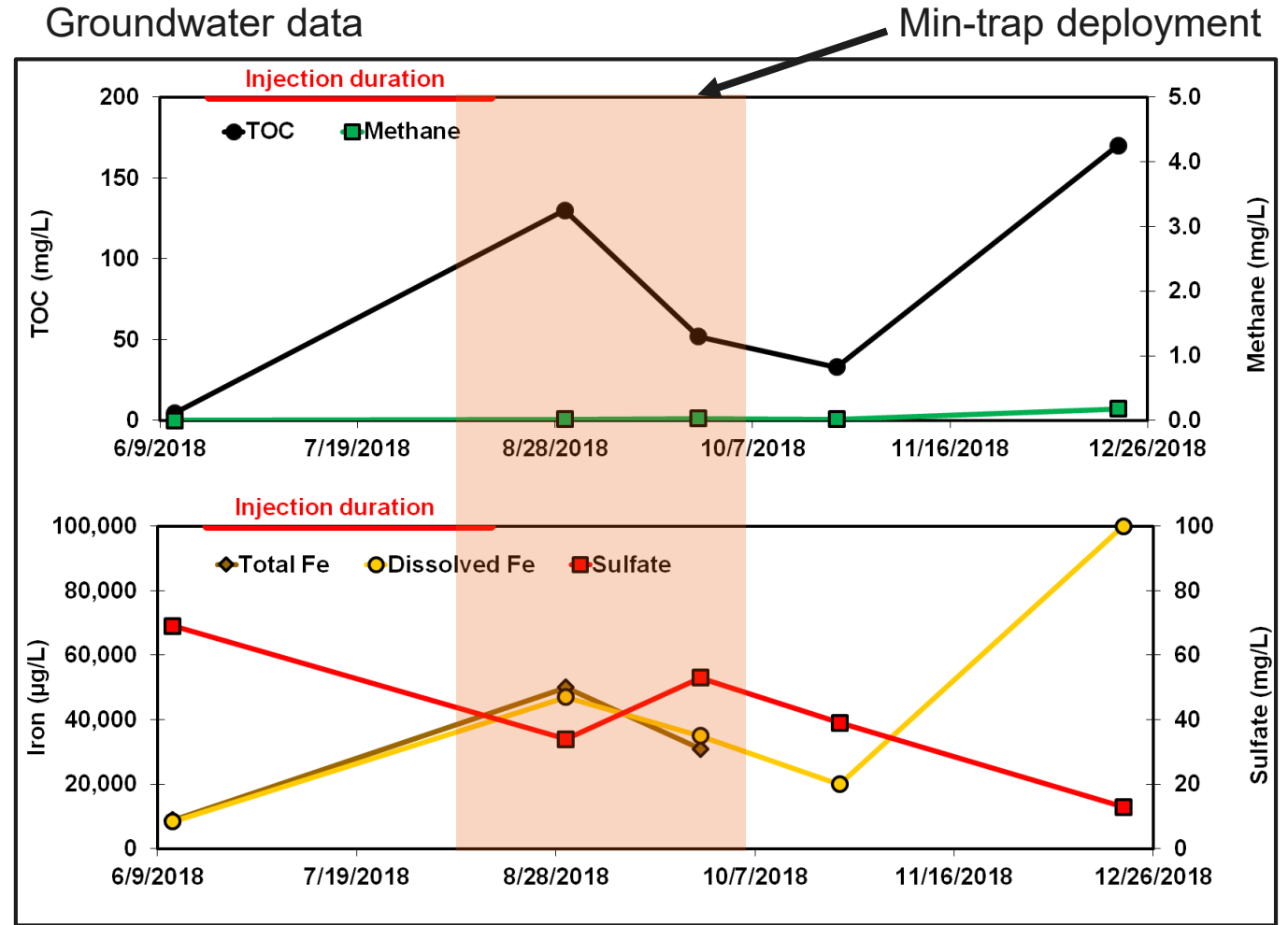


Post-Injection Performance



Iron: Solid iron is reduced
Sulfur: Mostly FeS, some FeS₂

Groundwater data



WAS Iron (mg/kg)	SAS Iron (mg/kg)	AV Sulfide (mg/kg)	CrE Sulfide (mg/kg)
Fe2+ = 330	Fe2+ = 300	240	120
Fe3+ = 0	Fe3+ = 30		

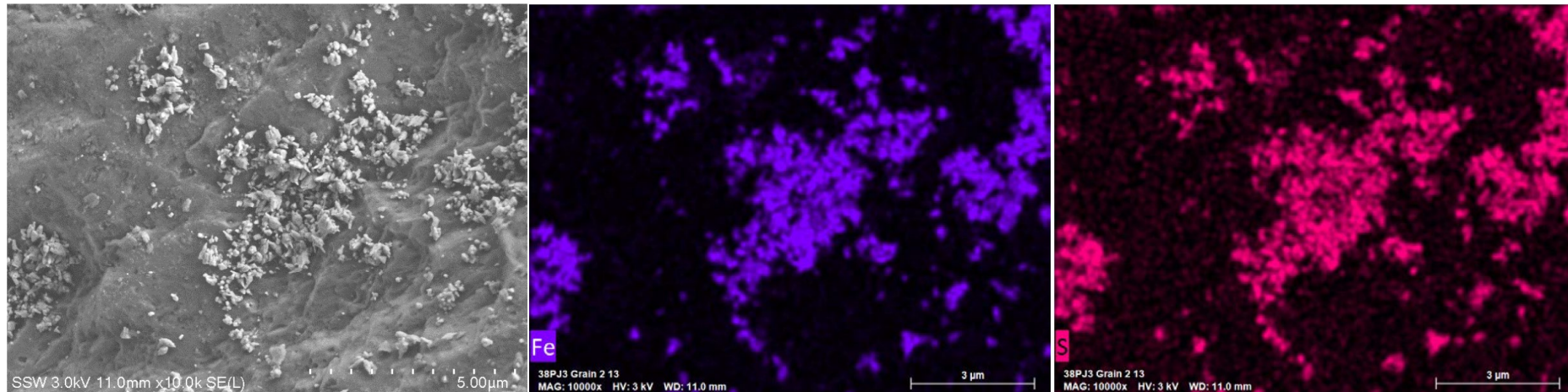


Post-Injection Performance: SEM with Energy Dispersive X-Ray Spectroscopy (EDS)

Backscatter micrograph

Iron

Sulfur



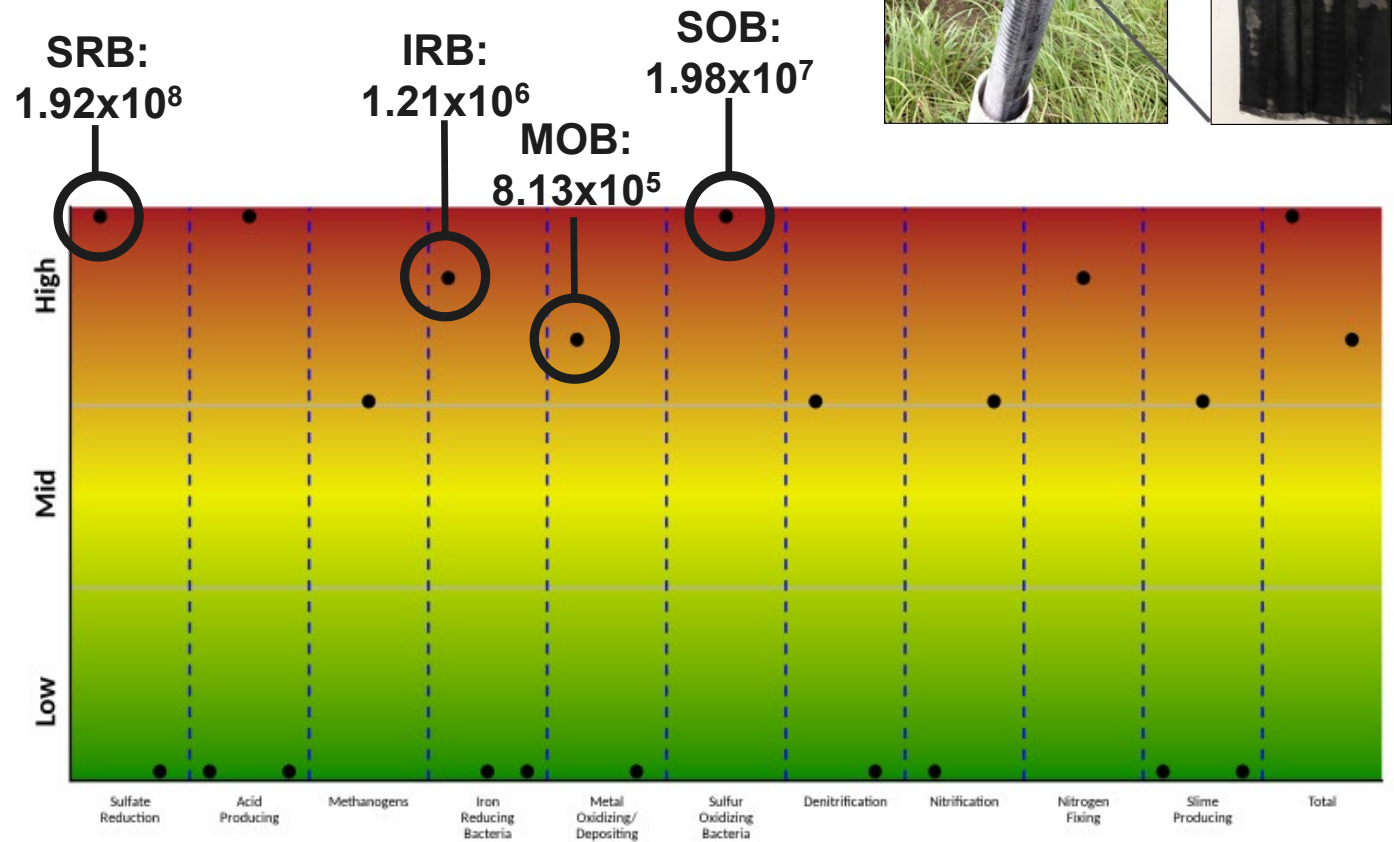
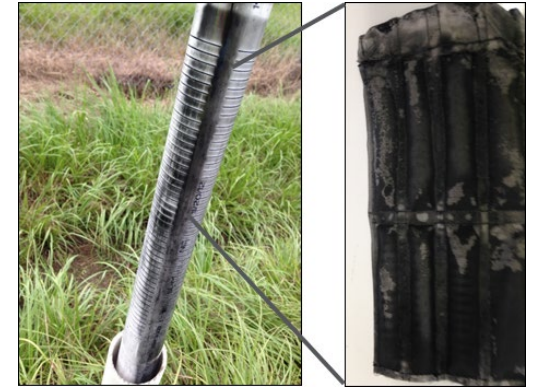
EDS spectra confirmed presence of iron and sulfur particles



Co-located iron and sulfur particles indicate that precipitates are iron-sulfur minerals

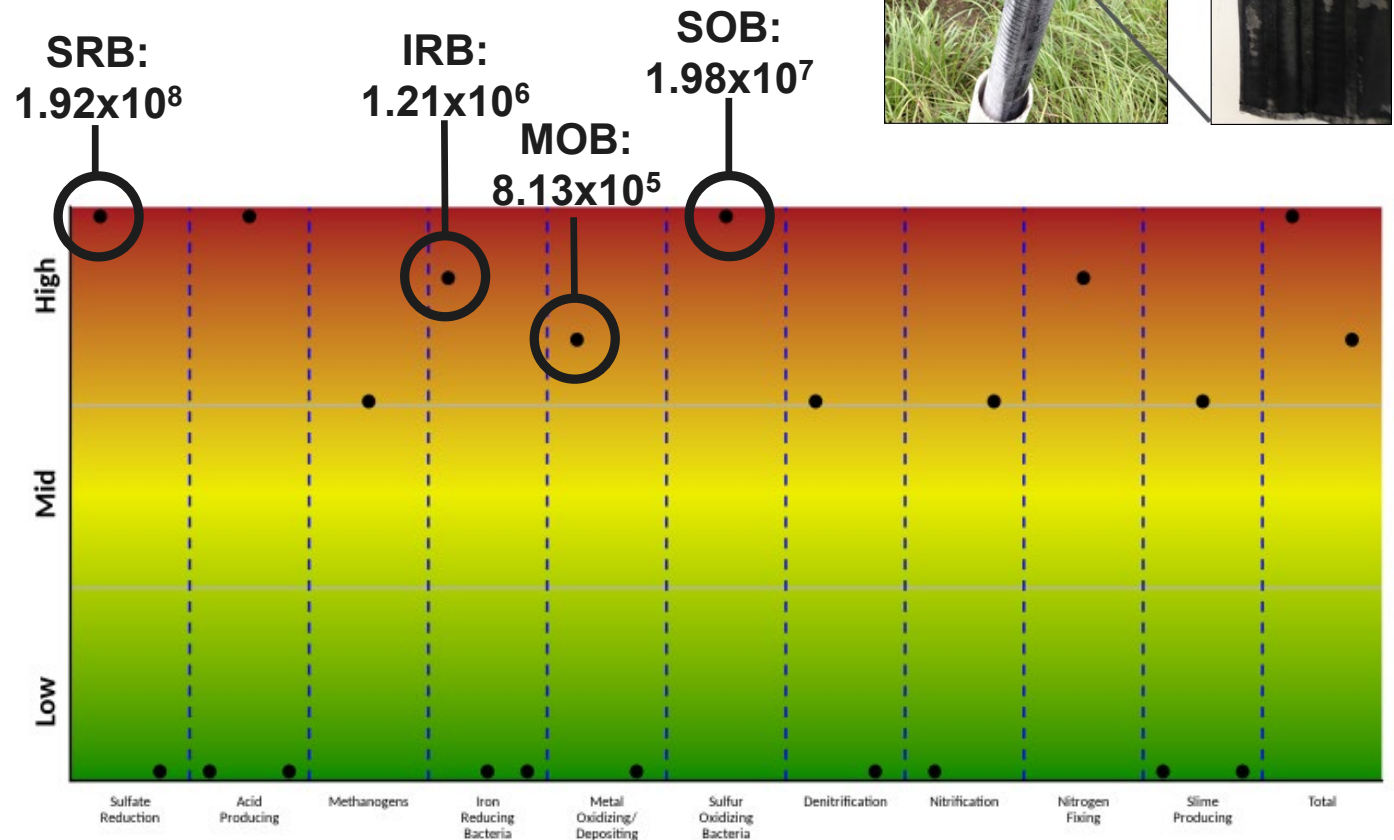
Post-Injection Performance: QuantArray-MIC

Sample Name	OBS-5
Sample Date	10/09/2018
<i>Microbial Induced Corrosion</i>	cells/g
Total Bacteria (EBAC)	7.74E+08
Total Archaea (ARC)	3.58E+05
Sulfate Reducing Bacteria (APS)	1.92E+08
Sulfate Reducing Archaea (SRA)	<1.00E+04
Methanogens (MGN)	1.69E+04
Acetogens (AGN)	<1.00E+04
Fermenters (FER)	3.11E+08
Iron Reducing Bacteria - Other (IRB)	1.21E+06
IRB <i>Geobacter</i> (IRG)	<1.00E+04
IRB <i>Shewanella</i> (IRS)	<1.00E+04
Iron Reducing Archaea (IRA)	<1.00E+04
Iron Oxidizers (FeOB)	8.13E+05
Manganese Oxidizing Bacteria (MnOB)	<1.00E+04
Sulfur Oxidizing Bacteria (SOB)	1.98E+07
Denitrifying Bacteria (nirK)	1.02E+04
Denitrifying Bacteria (nirS)	<1.00E+04
Ammonia Oxidizing Bacteria (AMO)	<1.00E+04
Nitrite Oxidizing Bacteria (NOR)	8.37E+04
Nitrogen Fixers (NIF)	5.57E+06
<i>Burkholderia cepacian</i> Exopolysaccharide (BCE)	<1.00E+04
<i>Deinococcus</i> spp. (DCS)	5.35E+04
<i>Meiothermus</i> spp. (MTS)	<1.00E+04
<i>Cladosporium</i> spp. CLAD	<1.00E+04
<i>CENSUS Targets</i>	
<i>Dehalobacter</i> spp. (DHBt)	2.42E+07
<i>Dehalobacter</i> DCM (DCM)	2.74E+05
Chloroform Reductase (CFR)	6.00E+03 (J)

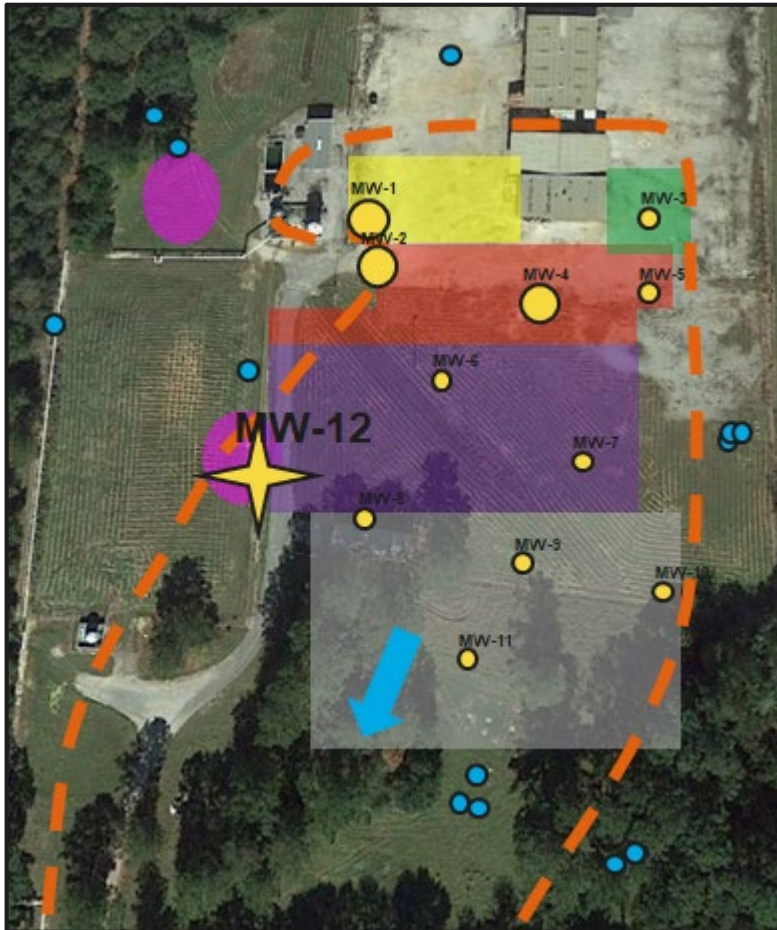


Post-Injection Performance: QuantArray-MIC

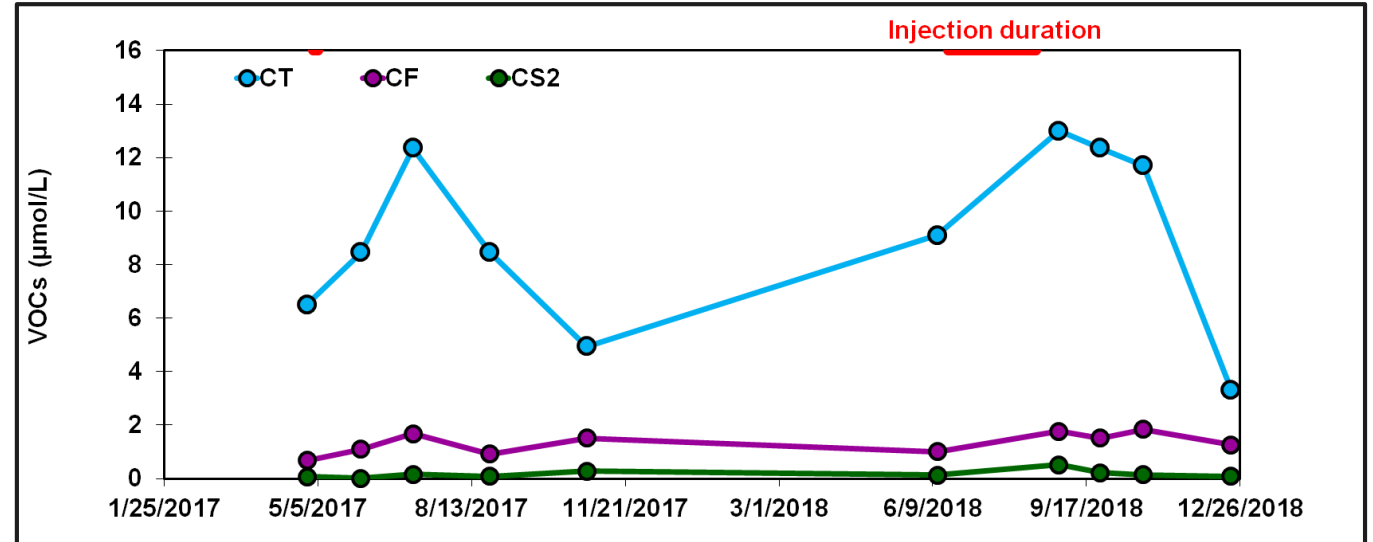
- Data provide insight on:
 - ✓ Geochemical conditions
 - ✓ Abundance of key microbial groups for degradation of CT/CF and formation of reactive minerals species
- Microbial analyses can be performed with Min-Trap samples
 - ✓ Media appropriate for sampling
- Min-Trap data comparable to corresponding groundwater data



Post-Injection Performance

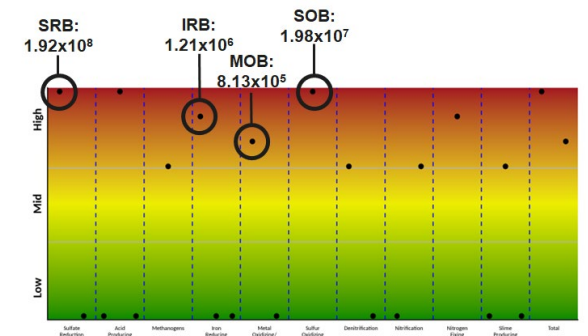
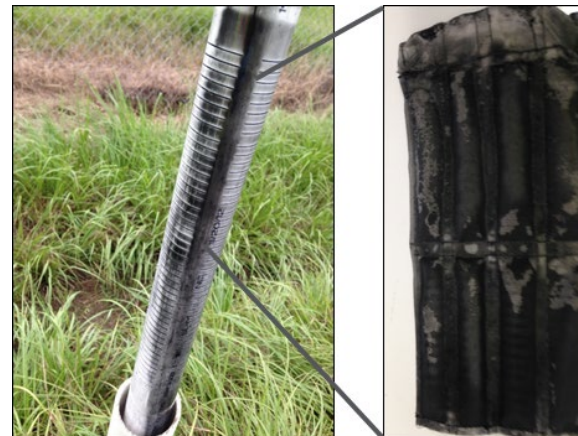
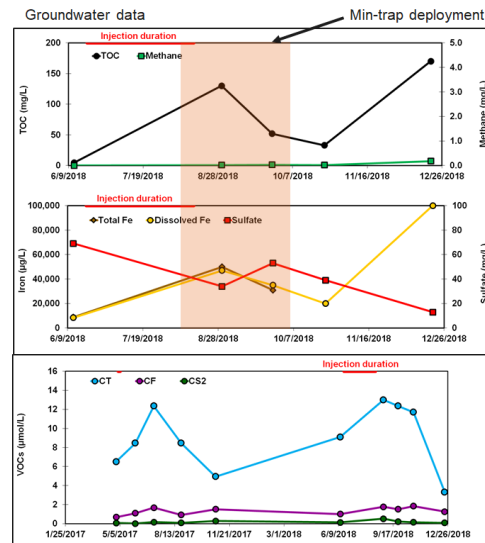
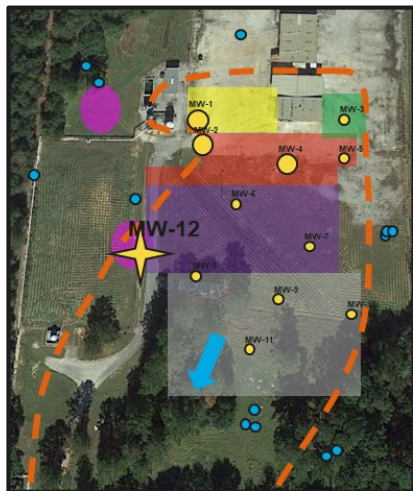


Groundwater data



Conclusions

- CT and CF were successfully treated via *in situ* EHC™ injections
- The selected remedial approach enhanced biotransformation, as well as abiotic and coupled microbial-abiotic transformation of CT and CF
- Remedial performance was evidenced through evaluation of geochemical conditions and CT/CF concentration trends, characterization of reactive iron sulfur minerals, and analysis of microbial community



Questions?



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Arcadis



Jennifer Tilton
Arcadis

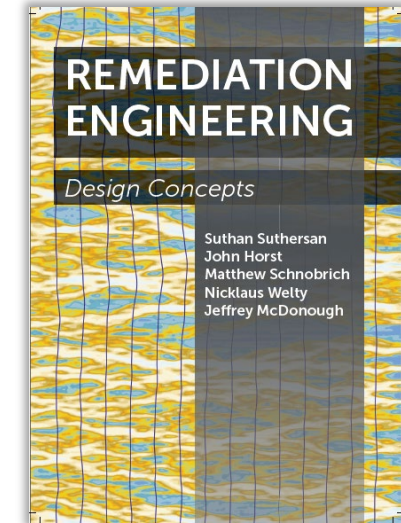
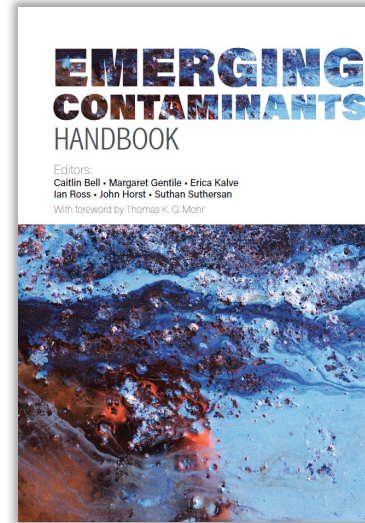


Dora Taggart,
Microbial Insights

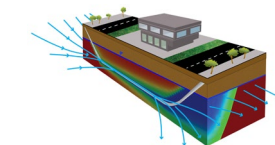


Kate Clark, PhD
Microbial Insights

Technical Knowledge



Innovation



HRX Well™
(Horizontal Treatment Well)

