

Transport of *Pseudonocardia* through Soil for Bioremediation of 1,4-Dioxane

David L. Freedman and Ángel A. Ramos-García

Clemson University

Department of Environmental Engineering & Earth Sciences

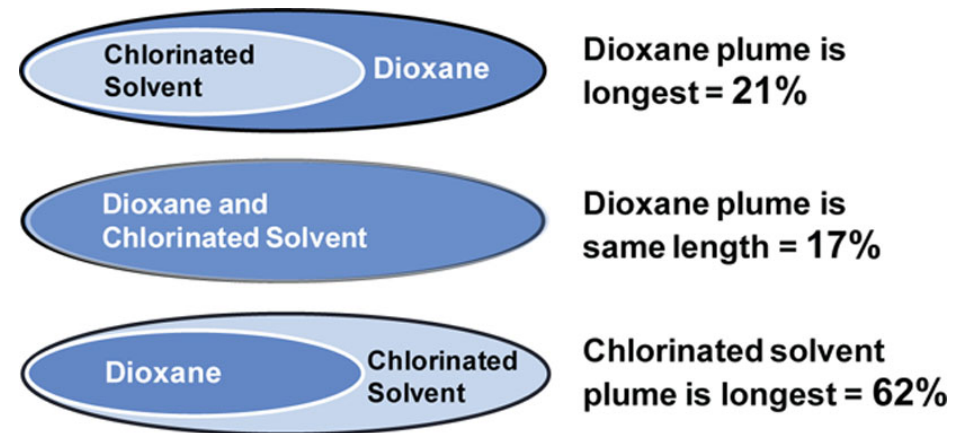
April 17, 2019



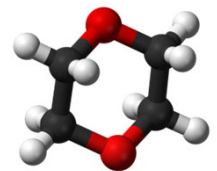
Overview

- Most 1,4-dioxane plumes seem to be shorter than expected
- Finding evidence of natural attenuation is challenging
- Some plumes will require active remediation
- Bioaugmentation may be required
- How well do 1,4-dioxane degrading cultures move through soil?

Adamson et al. (2014):



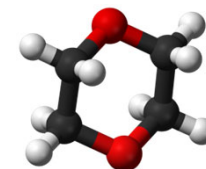
n = 103 sites where dioxane and chlorinated solvents co-occur



Overview

1,4-Dioxane Aerobic Cultures for Bioaugmentation

- Use 1,4-dioxane as a growth substrate
 - Best known is *Pseudonocardia dioxivorans* CB1190
 - Isolated from activated sludge
 - Tendency to clump when grown in the lab; raises questions about movement through soil for bioaugmentation applications
- Cometabolize 1,4-D following growth on a primary substrate
 - Various isolates that grow on various substrate (methane, propane, butane, etc.)
 - Less propensity to clump?

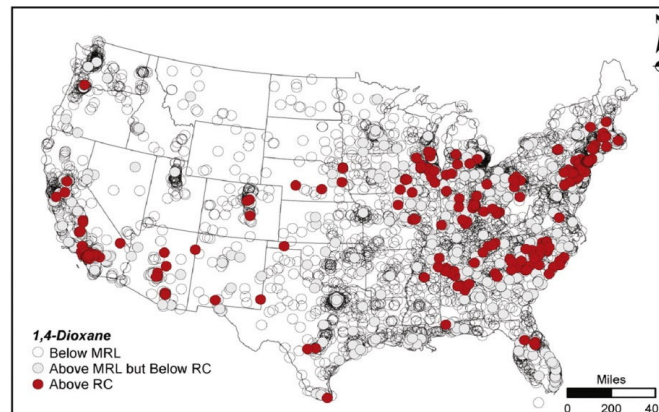


Objectives

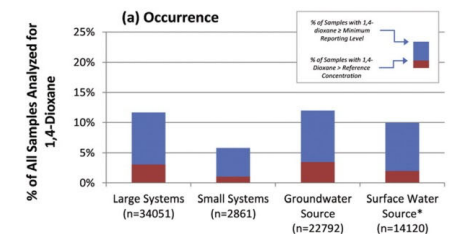
- Enrich and isolate 1,4-D degraders from 2 aquifers
 - Industrial site in the southeastern US
 - Industrial site in Europe
- Compare one of the aquifer isolates to CB1190 in movement through porous media
 - Sand
 - Silt

Adamson et al. (2017):

1,4-Dioxane Occurrence in 4864 Public Water Systems Included in UCMR3



1,4-Dioxane detected in 21% of public water systems but detection rates declined over time

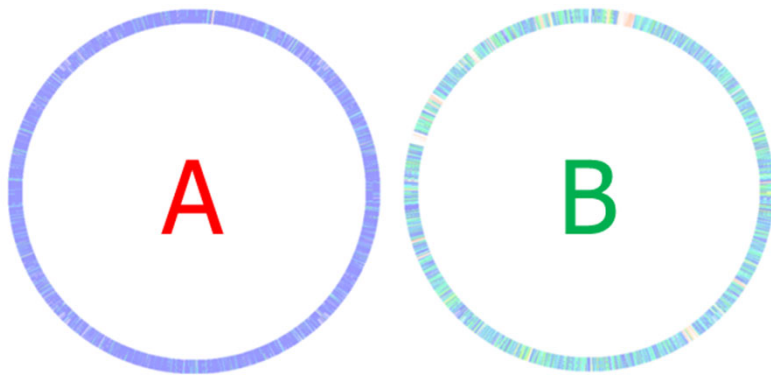
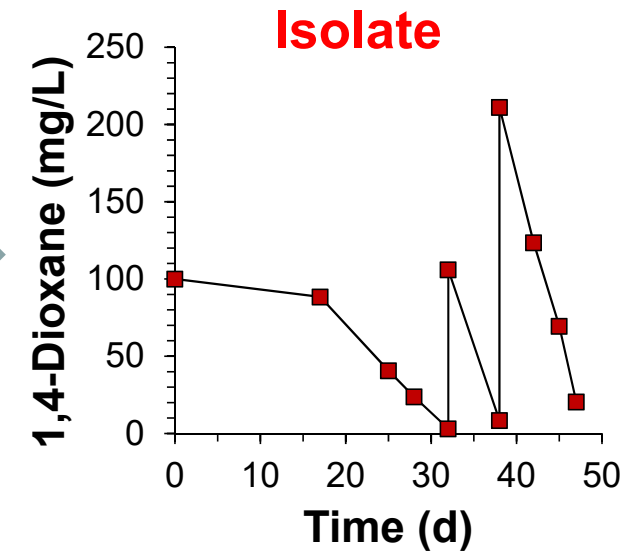
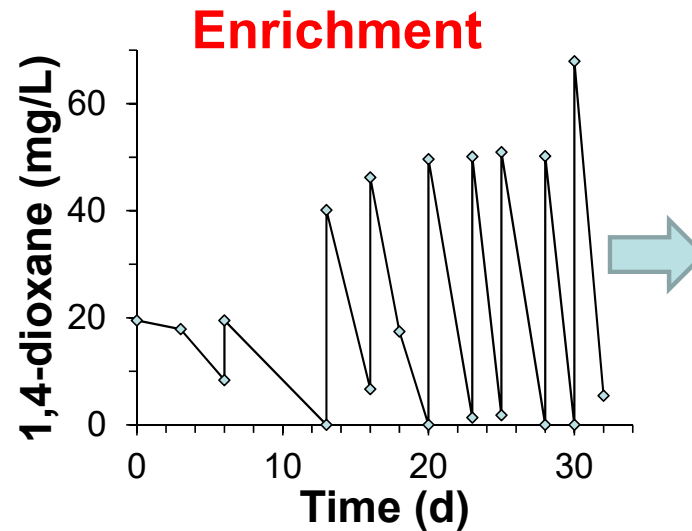
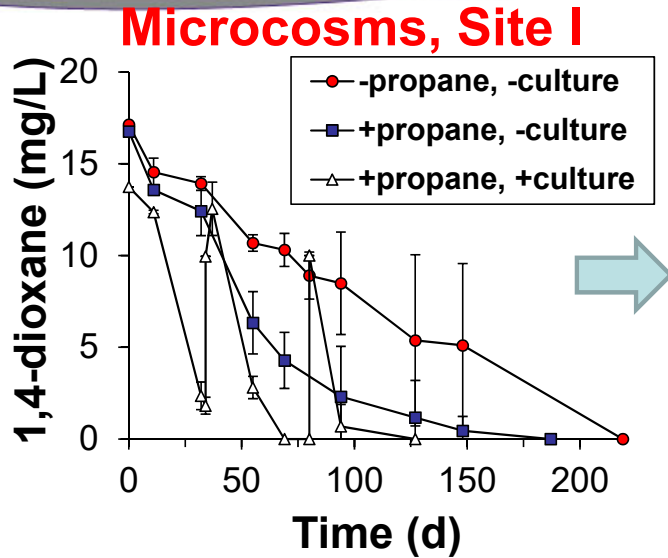


Exposure not solely related to groundwater-based conceptual model for 1,4-dioxane releases

1,4-D Isolates

- Microcosms prepared with soil (20 g) + GW (50 mL)
 - Treatments evaluated natural attenuation, propane addition, and propane + propanotrophic culture
- Enriched microcosms with only 1,4-D and transfers to ammonium mineral salts medium (AMSM)
- Isolates obtained by repeated streaking on Noble agar + AMSM plates with 50 mg/L 1,4-dioxane
- Whole genome sequencing performed at the UNC-Chapel Hill Microbial Core Facility
- *Pseudonocardia dioxanivorans* strain CB1190 provided courtesy of Dr. Shaily Mahendra, UCLA

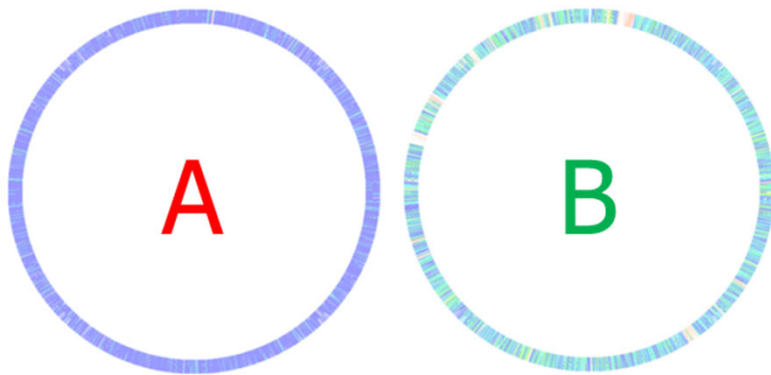
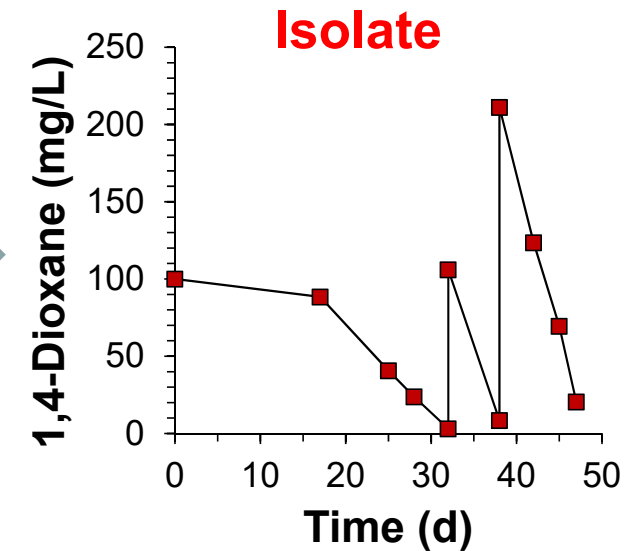
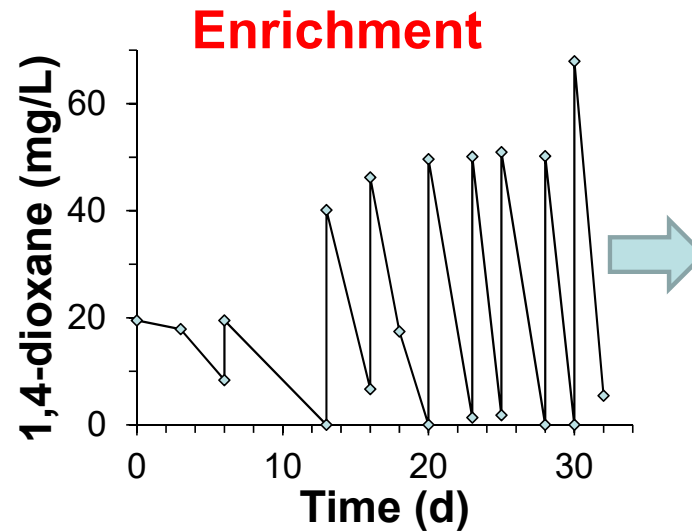
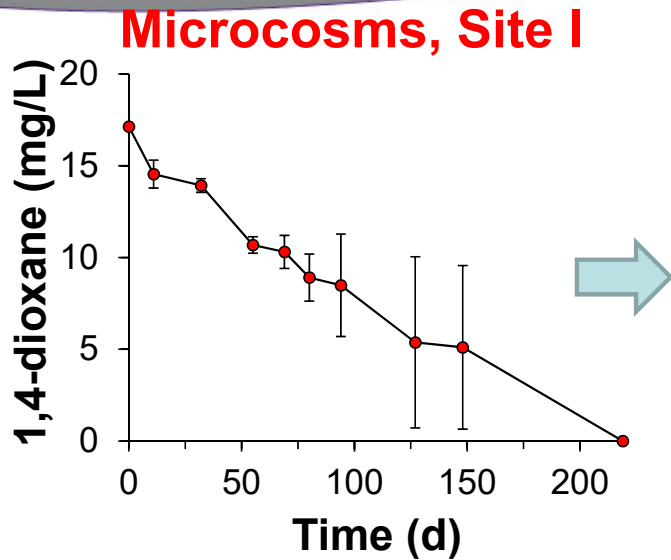
Isolate #1



- **16S rRNA sequence: same as CB1190**
- **Gene-to-gene comparison**
 - A = Positive control, CB1190 from literature vs. CB1190 at Clemson → nearly identical**
 - B = CB1190 vs. isolate #1 → different strains; isolate named “BERK-1”**

Ramos-Garcia, et al. 2018. Draft genome sequence of the 1,4-dioxane-degrading bacterium *Pseudonocardia dioxanivorans* BERK-1. *Genome Announcements* 6 (14), e00207-18.

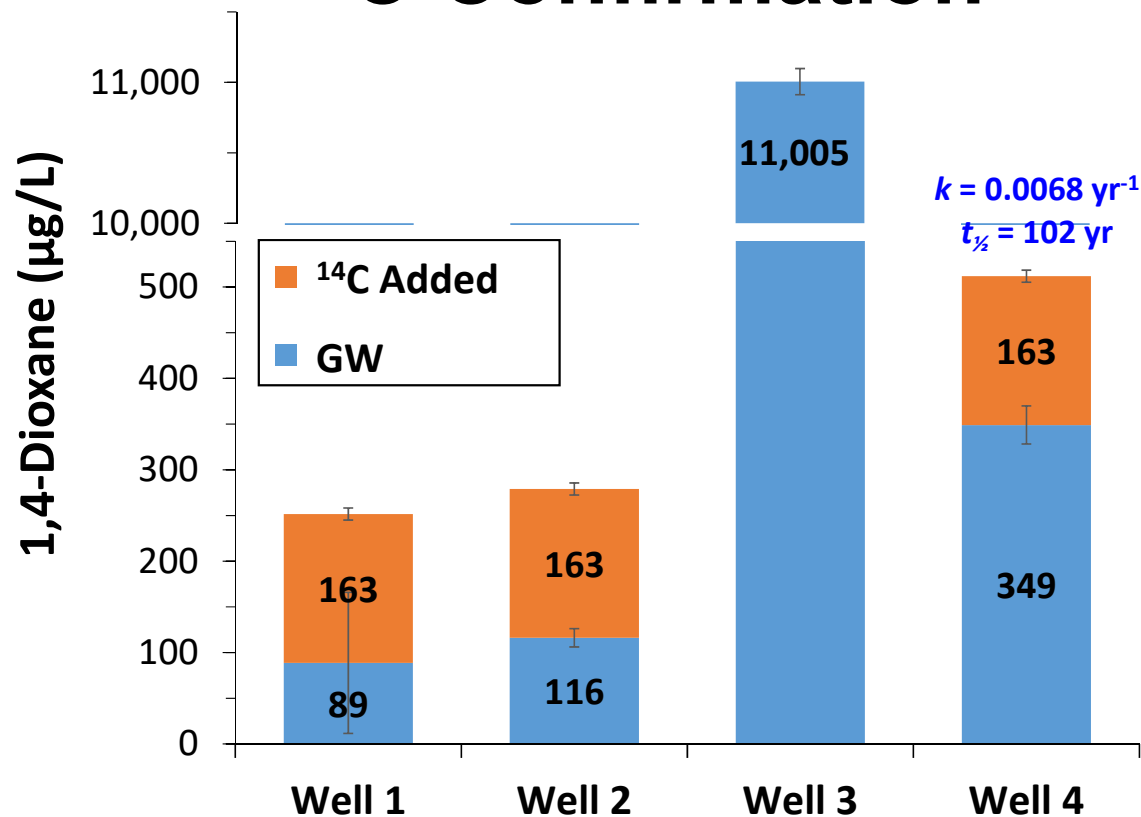
Isolate #1



- **16S rRNA sequence: same as CB1190**
- **Gene-to-gene comparison**
 - A = Positive control, CB1190 from literature vs. CB1190 at Clemson → nearly identical**
 - B = CB1190 vs. isolate #1 → different strains; isolate named “BERK-1”**

Ramos-Garcia, et al. 2018. Draft genome sequence of the 1,4-dioxane-degrading bacterium *Pseudonocardia dioxanivorans* BERK-1. *Genome Announcements* 6 (14), e00207-18.

¹⁴C Confirmation

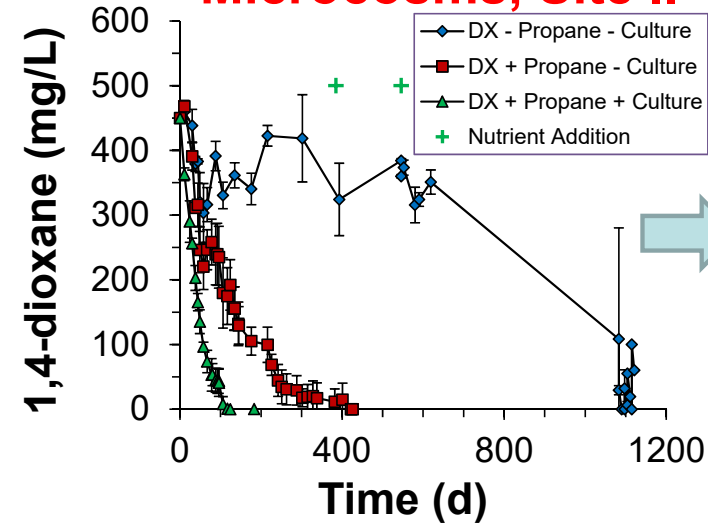


New samples of GW from site I collected

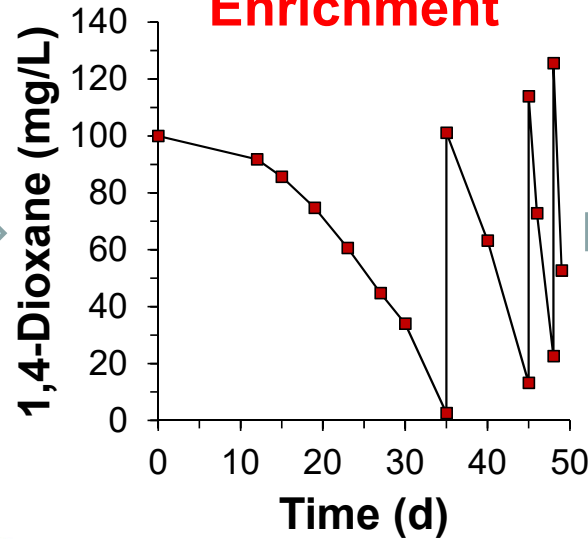
- Part of ESTCP Project ER-201730: *Development of a Quantitative Framework for Evaluating Natural Attenuation of 1,1,1-TCA, 1,1-DCA, 1,1-DCE, and 1,4-Dioxane in Groundwater*
- Biodegradation of 1,4-D observed in samples from one well
- Low rate constant is consistent with a slow rate of degradation in original microcosms
- VOC levels are low, non-inhibitory

Isolate #2

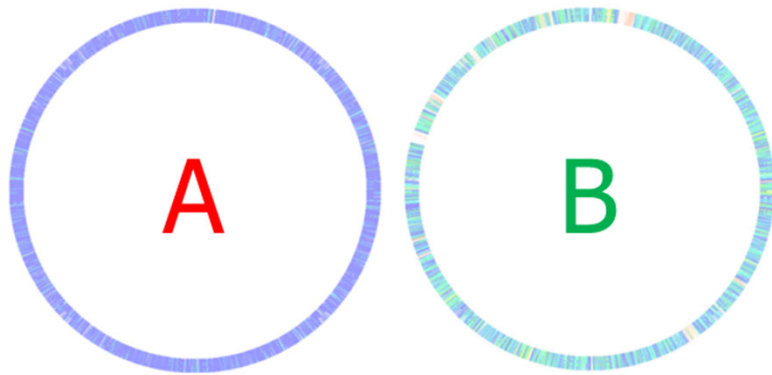
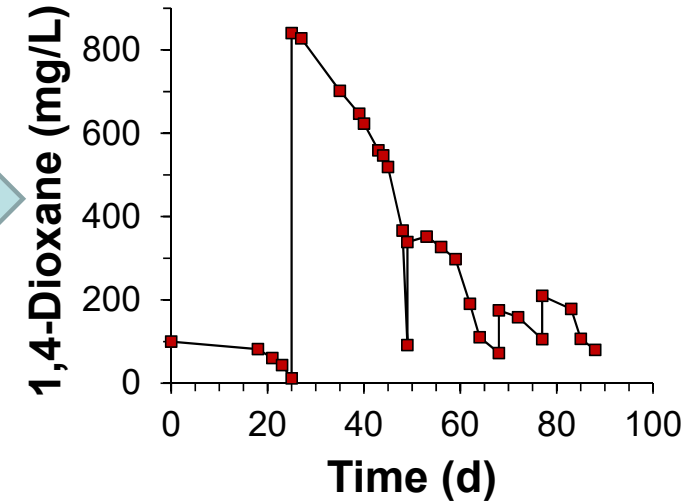
Microcosms, Site II



Enrichment



Isolate



- 16S rRNA sequence: same as CB1190
- Gene-to-gene comparison:
 - A = BERK-1 versus EUR-1 → nearly identical; same strain
 - B = EUR-1 versus CB1190 → different strains

Isolates



CB1190



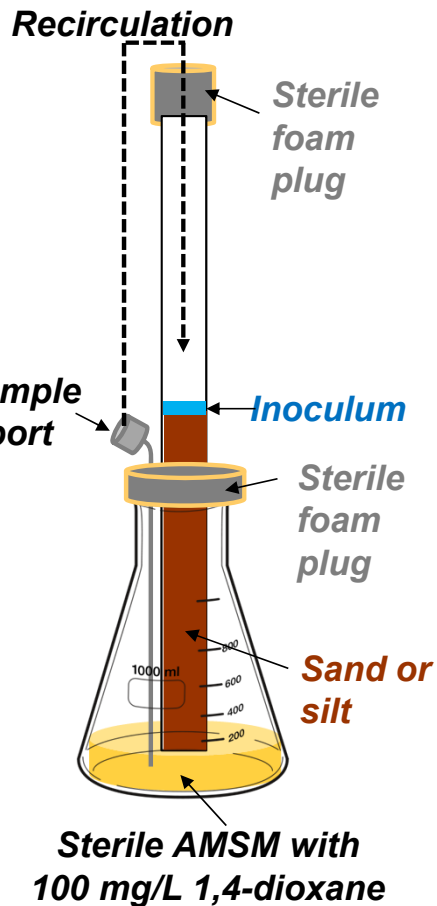
**BERK-1
from site II**



**BERK-1
from site I**

- Significantly greater clumping and attachment by CB1190 versus BERK-1
- Is this related to the origin of the isolates, i.e., activated sludge vs. aquifers?
- Hypothesis: BERK-1 will move through porous medium at a higher rate since it is less “sticky”

Movement of Isolates

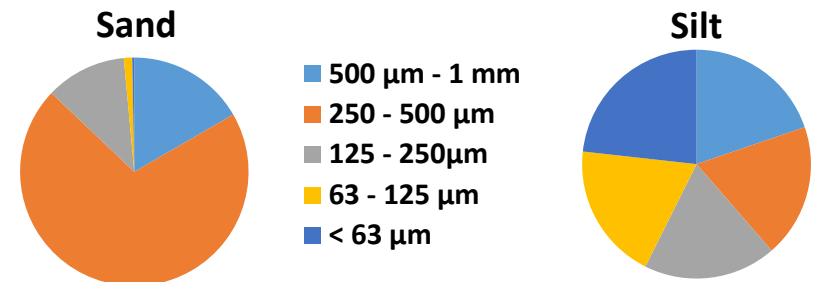


- Glass tube: 0.87" inner diameter x 11.8" long; 5" filled with sand or silt
- Glass wool plug at base retained the sand or silt

Porosity:

sand ~0.4

silt ~0.3

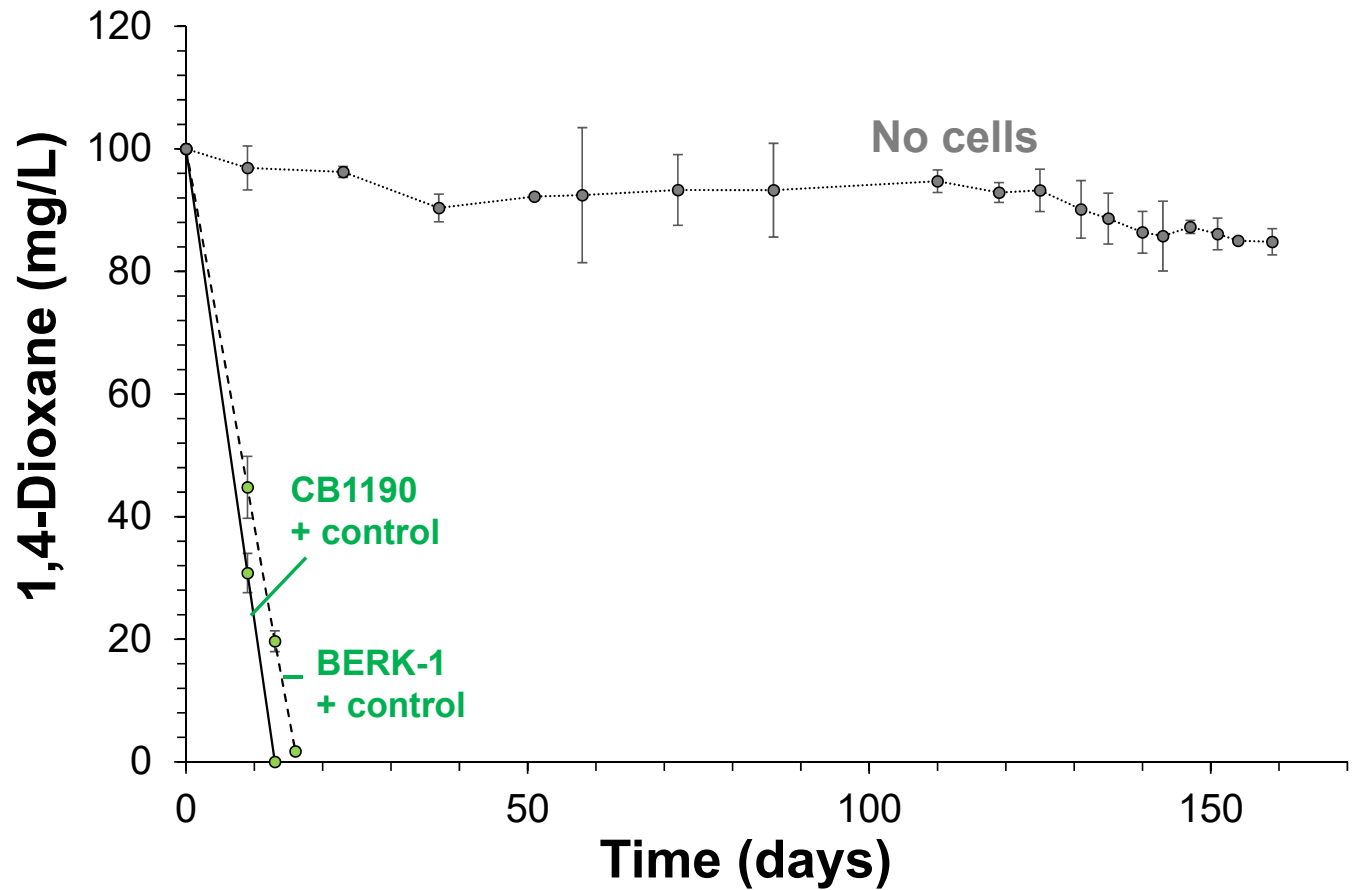
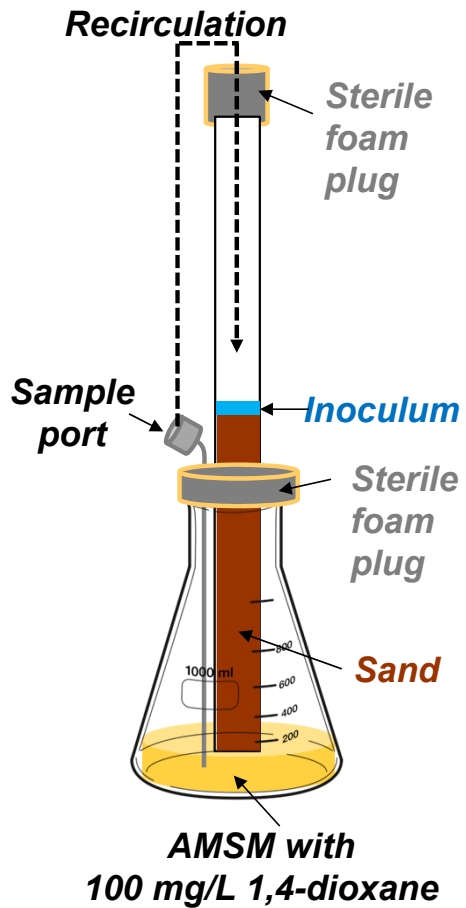


Inocula:

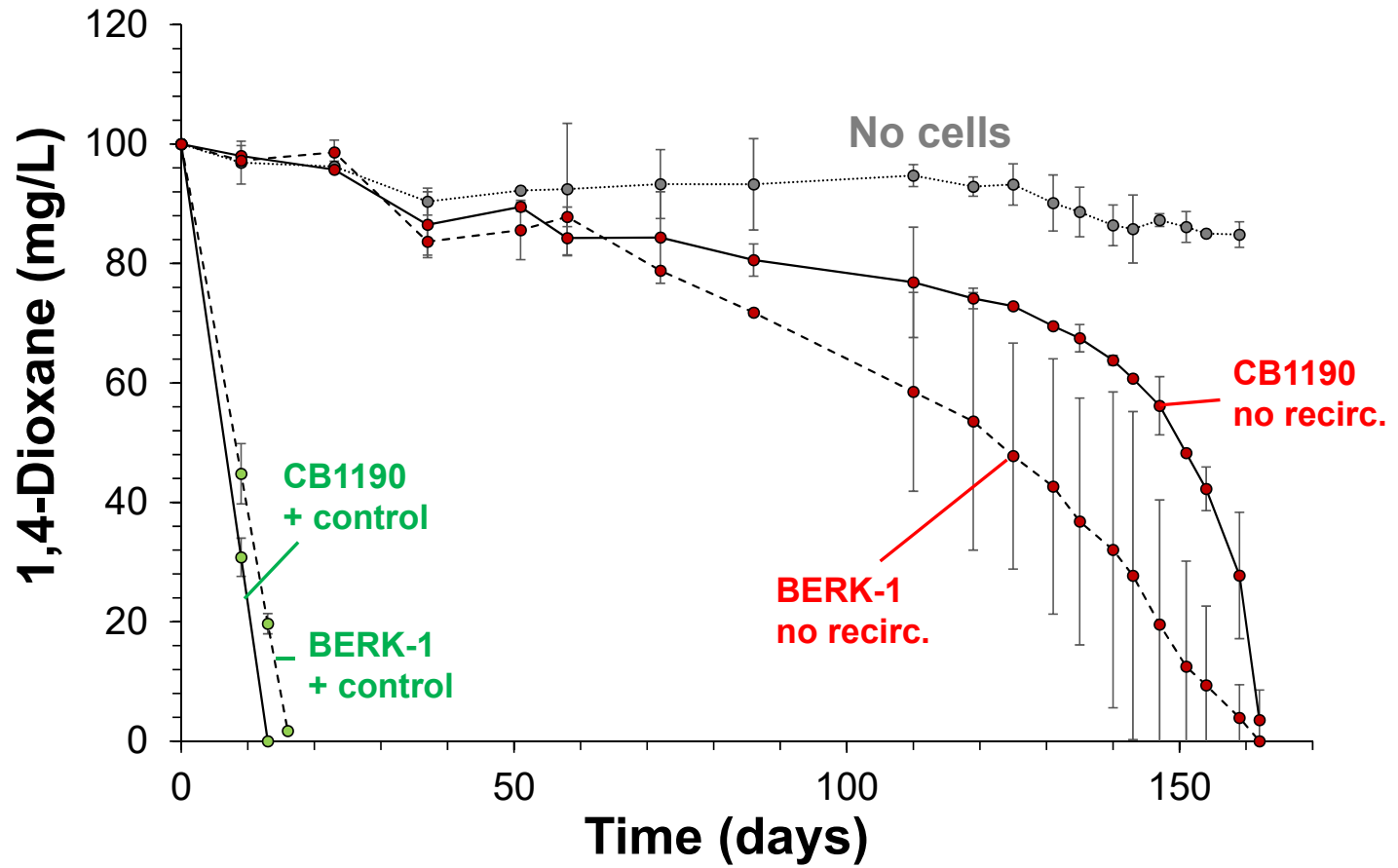
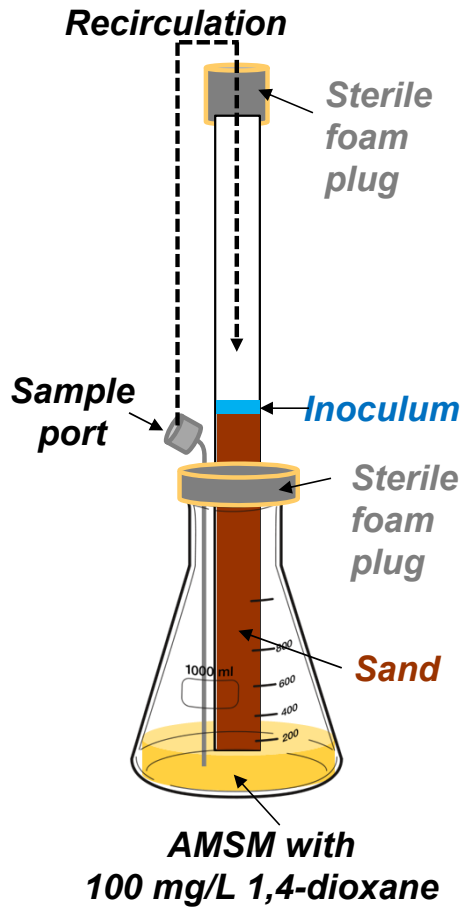
- 6 mL of CB1190 or BERK-1 grown on 1,4-dioxane;
- ~0.13 mg protein/mL ($\sim 10^8$ cells/mL)

- Positive control: directly inoculate the MSM

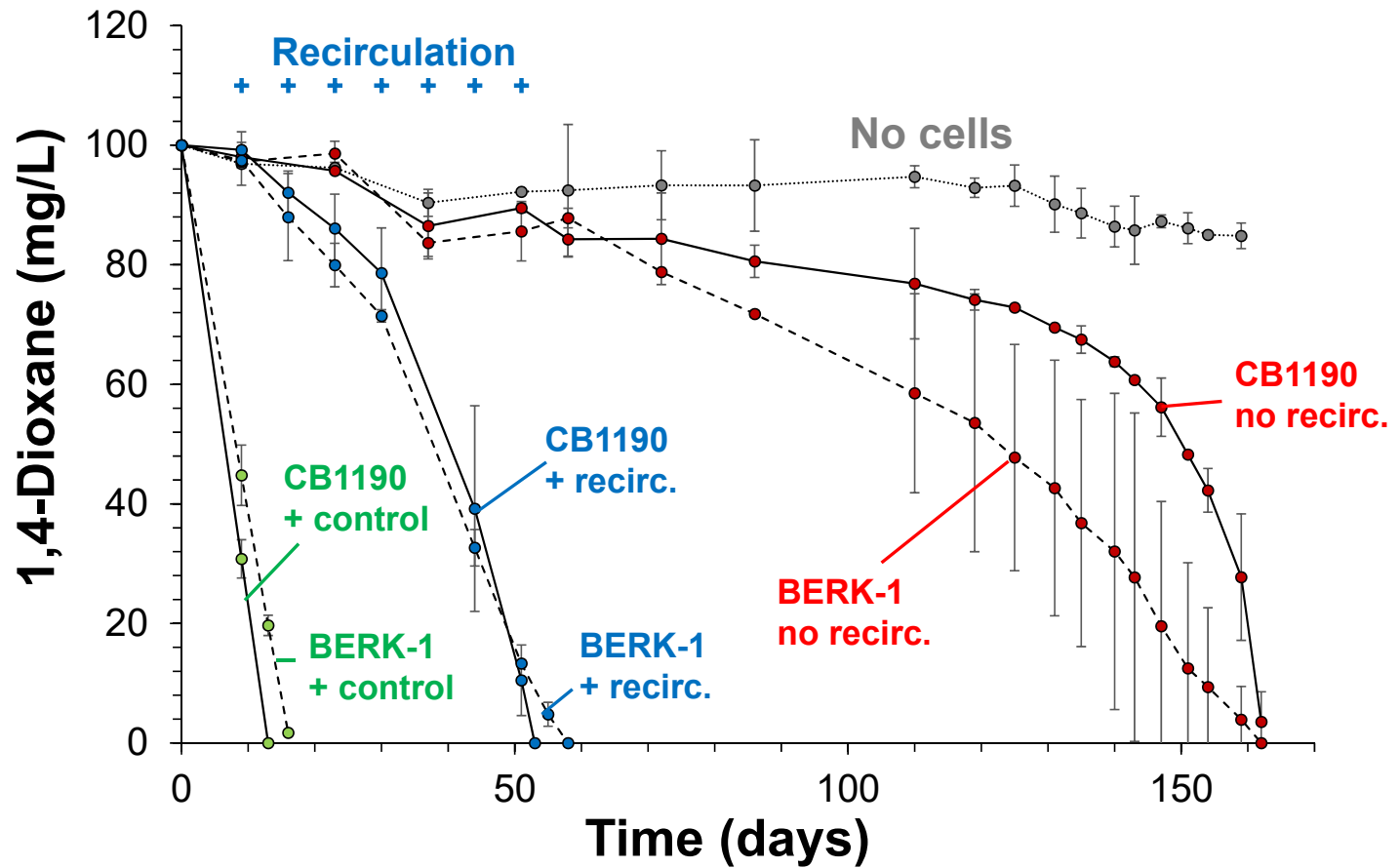
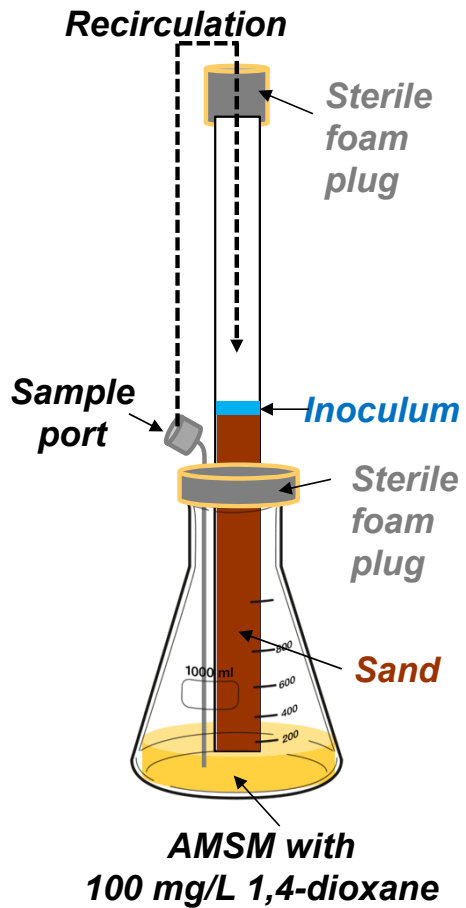
Movement of Isolates: Sand



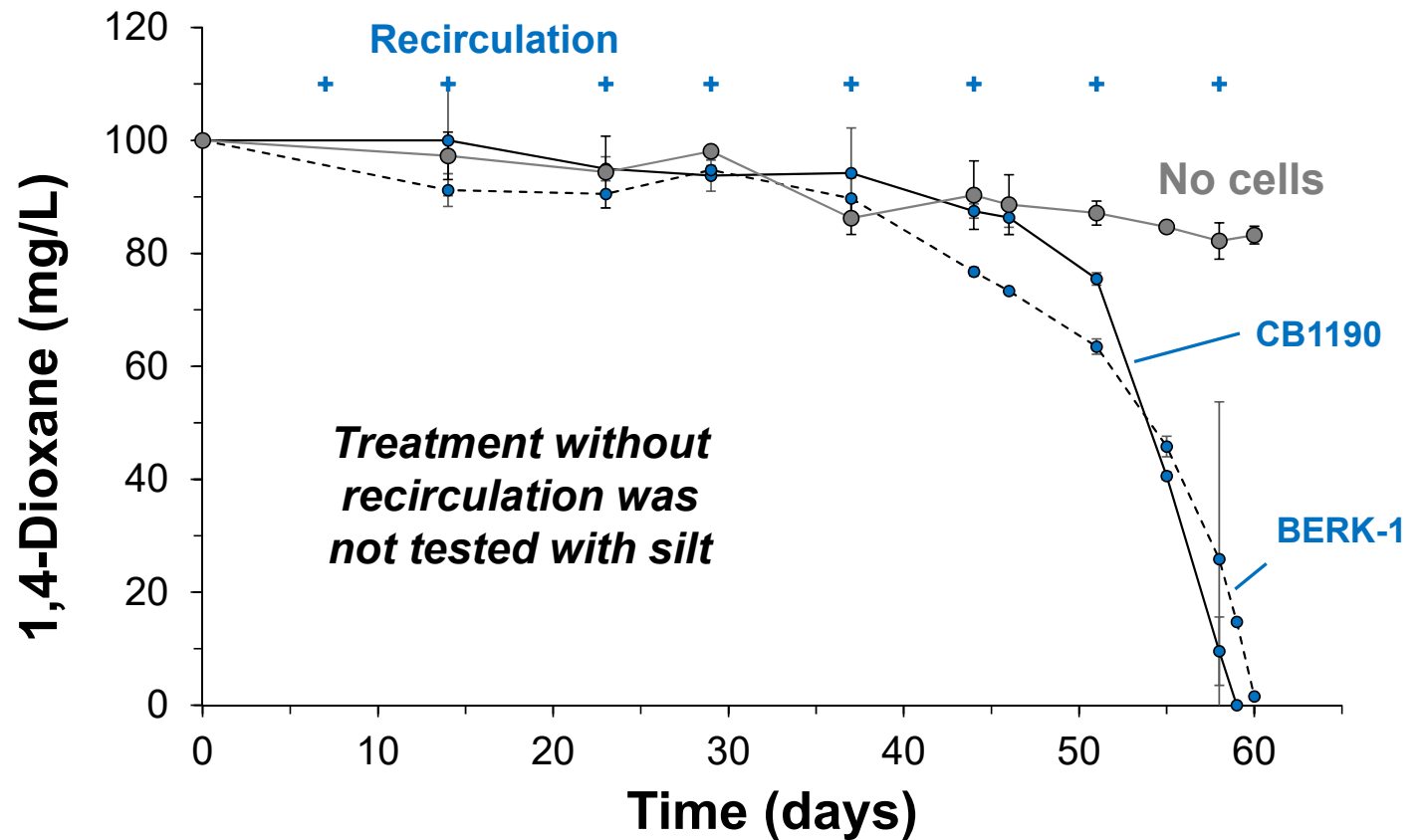
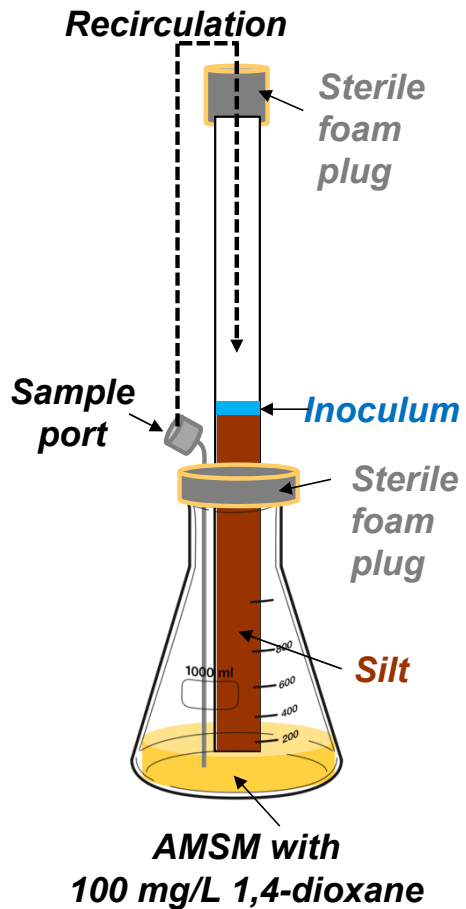
Movement of Isolates: Sand



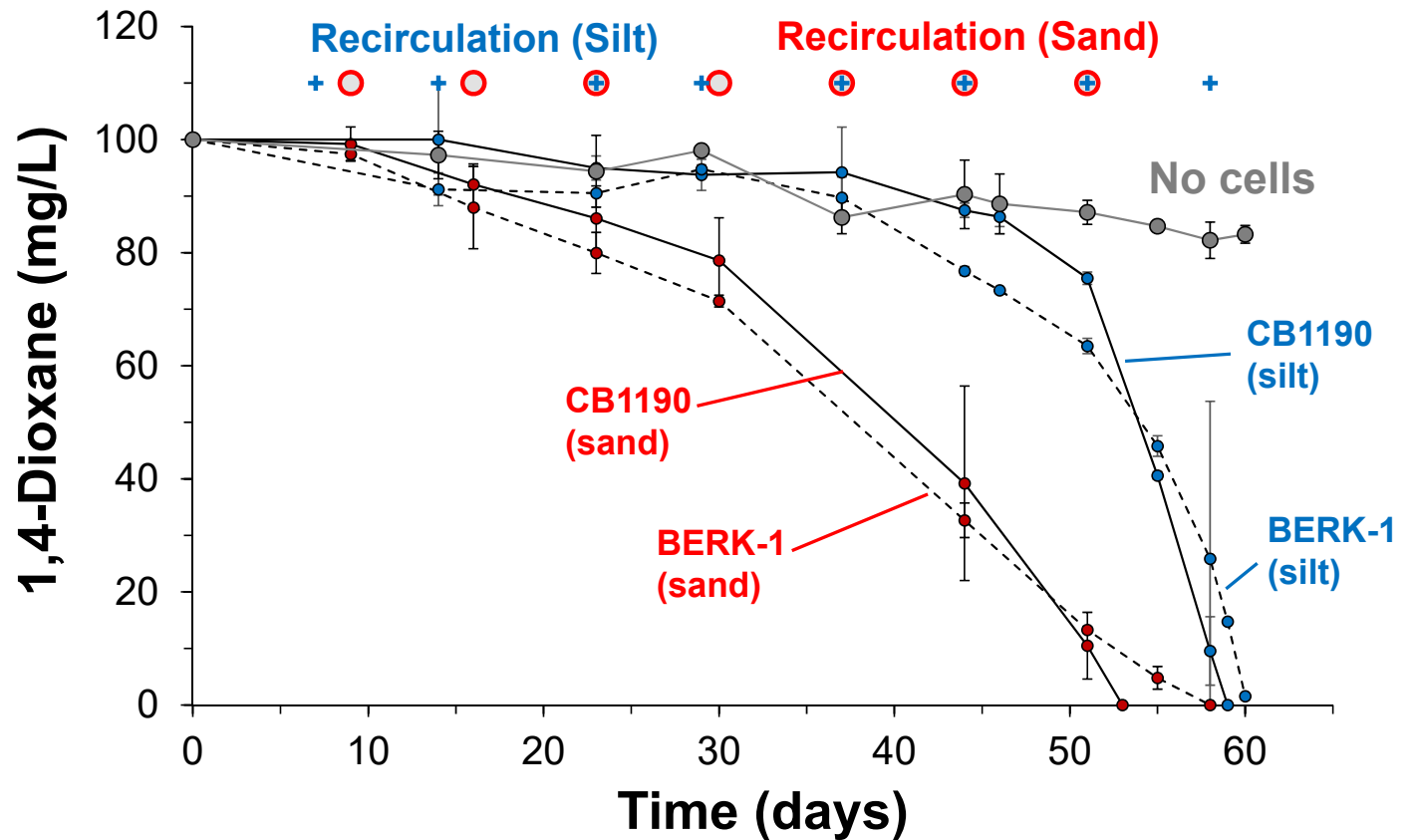
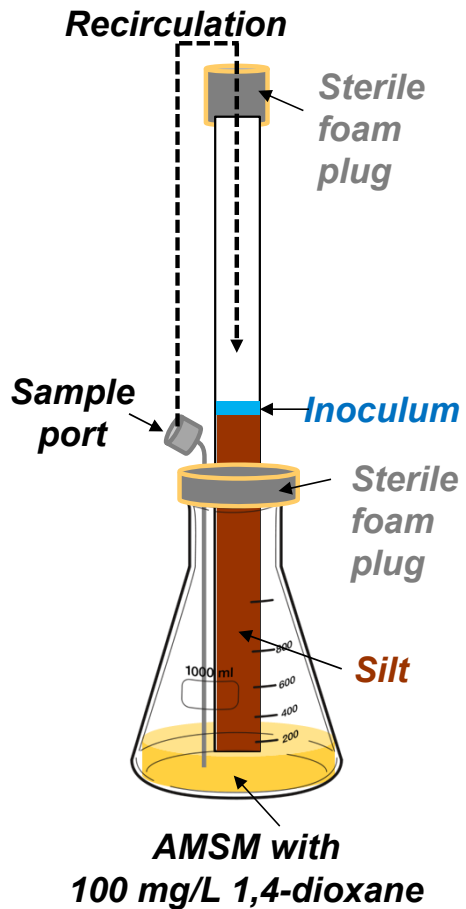
Movement of Isolates: Sand



Movement of Isolates: Silt



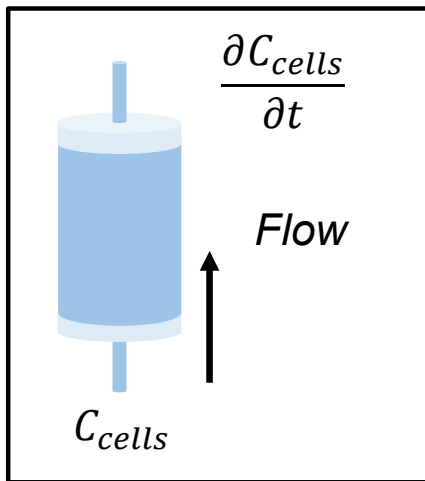
Movement of Isolates: Sand vs. Silt



Conclusions

- 2 isolates obtained from aquifers; same genus and species as *Pseudonocardia dioxivorans*, different strain
- Likely presence of BERK-1 at southeastern US site confirmed with a ^{14}C assay; low 1st order rate consistent with microcosms
 - *In situ* 1,4-D degraders probably limited by low oxygen and/or nutrients
- CB-1190 initially moved more slowly than BERK-1 through sand and silt, but made up for this by a higher rate of degradation
- Clumping behavior of cultures grown on rich medium is not necessarily a good predictor of transport *in situ*
 - Transport may be facilitated by conditioning the cultures, e.g., starvation response to reduce size and clumping
- Need to perform continuous flow column tests and develop a predictive model for microbial transport

Modeling Approach for Transport of Microbes through Soil



Perform column studies to determine parameters

$$\begin{aligned}
 & \text{Microbes} \quad \text{Mechanical Dispersion} \quad \text{Molecular Diffusion} \quad \text{Darcy's velocity} \\
 & \frac{\partial X}{\partial t} - (D_{D,X} + D_{F,X}) \frac{\partial^2 X}{\partial x^2} + (v_P) \frac{\partial X}{\partial x} + \frac{\partial X_{S'}}{\partial t} \\
 & = \\
 & \text{Specific Growth Rate (Monod)} \quad \mu_X \cdot (X + X_{S'}) \quad \text{Attachment and Detachment}
 \end{aligned}$$

Questions?

