

Development and Application of a Rapid, User-Friendly and Inexpensive Method to Detect *Dehalococcoides* spp. Genes in Groundwater

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Introduction

Technical Objective:

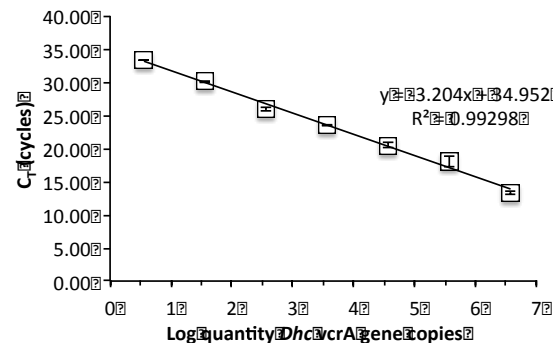
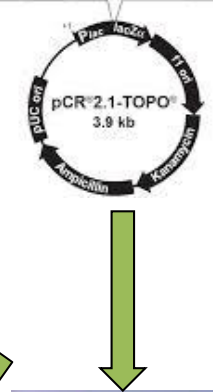
To develop rapid, sensitive and specific methods to quantify *Dehalococcoides* spp. genes from groundwater samples without DNA extraction

Key Components:

- ✓ Use loop mediated isothermal amplification (LAMP) as a technique for molecular detection
- ✓ Carry out validation with bioaugmentation cultures & groundwater samples
- ✓ Develop a LAMP based method which is low cost and easy to use
- ✓ Explore the development of field deployable approaches



- [illegible]

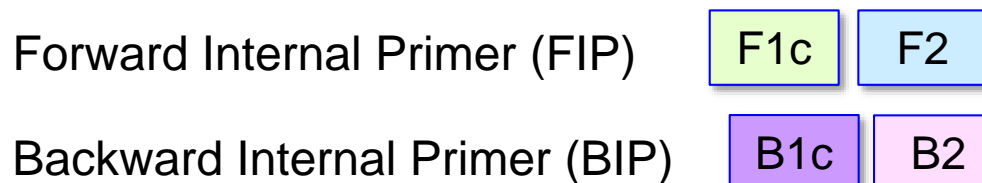
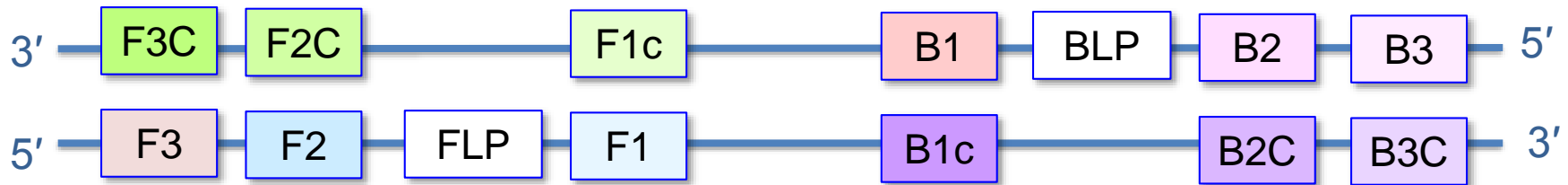




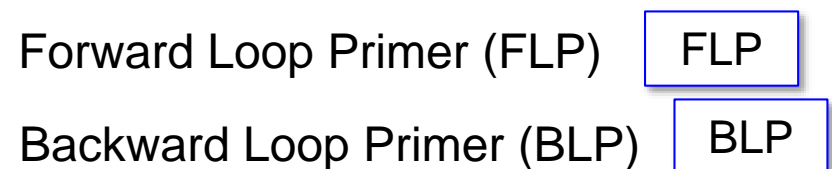
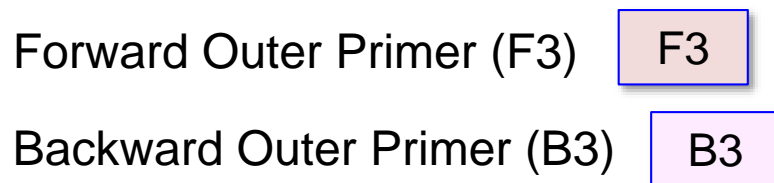
Introduction:

Loop Mediated Isothermal Amplification (LAMP)

Amplification creates stem loop structures with several inverted repeats of the target & cauliflower like structures with multiple loops



having both sense and antisense sequence helps in the formation of a loop





Introduction:

Applicability of LAMP for Development of Field Deployable Kits

- Amplification occurs at one temperature (**isothermal**), thermal cycler may not be needed
- **High amplification efficiency**, DNA is amplified 10^9 - 10^{10} times in 15 – 60 min.
- **Direct amplification** without DNA extraction for water samples
- Less sensitive to substances that inhibit PCR
- Six primers make LAMP **highly specific**
- Because of these advantages LAMP has been widely applied as a **point of care molecular diagnostic tool** for detection of a variety of templates

Step 1: Biomass Extraction

Direct cell extractions from groundwater



Step 2: Detection of template

Visual detection with dsDNA binding dyes



Detection on real time thermal cyclers



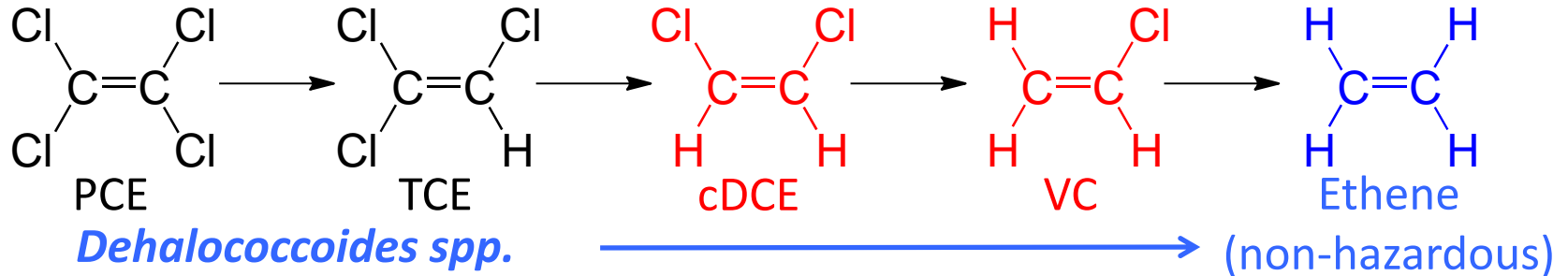
Detection on proprietary microfluidic platforms e.g. Gene-Z





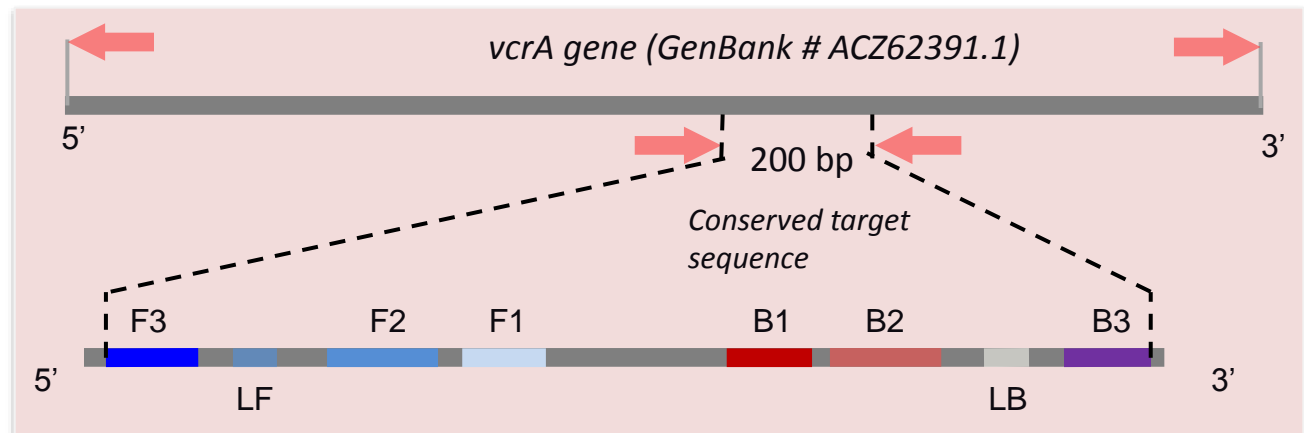
LAMP with Rdase genes

Primer Development



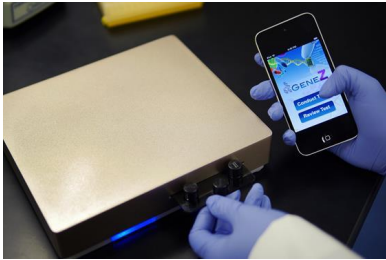
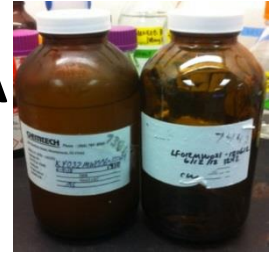
- Survival & activity is typically determined using qPCR targeting 16S rRNA or functional genes (RDase genes)
- Three LAMP primer sets specific to the *tceA*, *vcrA*, and *bvcA* genes were designed using Primer Explorer v4

LAMP primer designing software
PrimerExplorer



Three Study Questions

Q1. Is **LAMP comparable to qPCR** for DNA extracted from **cultures** or **groundwater**?



Q2. Is **LAMP with Gene-Z** comparable to **qPCR with a real time thermal cycler**?

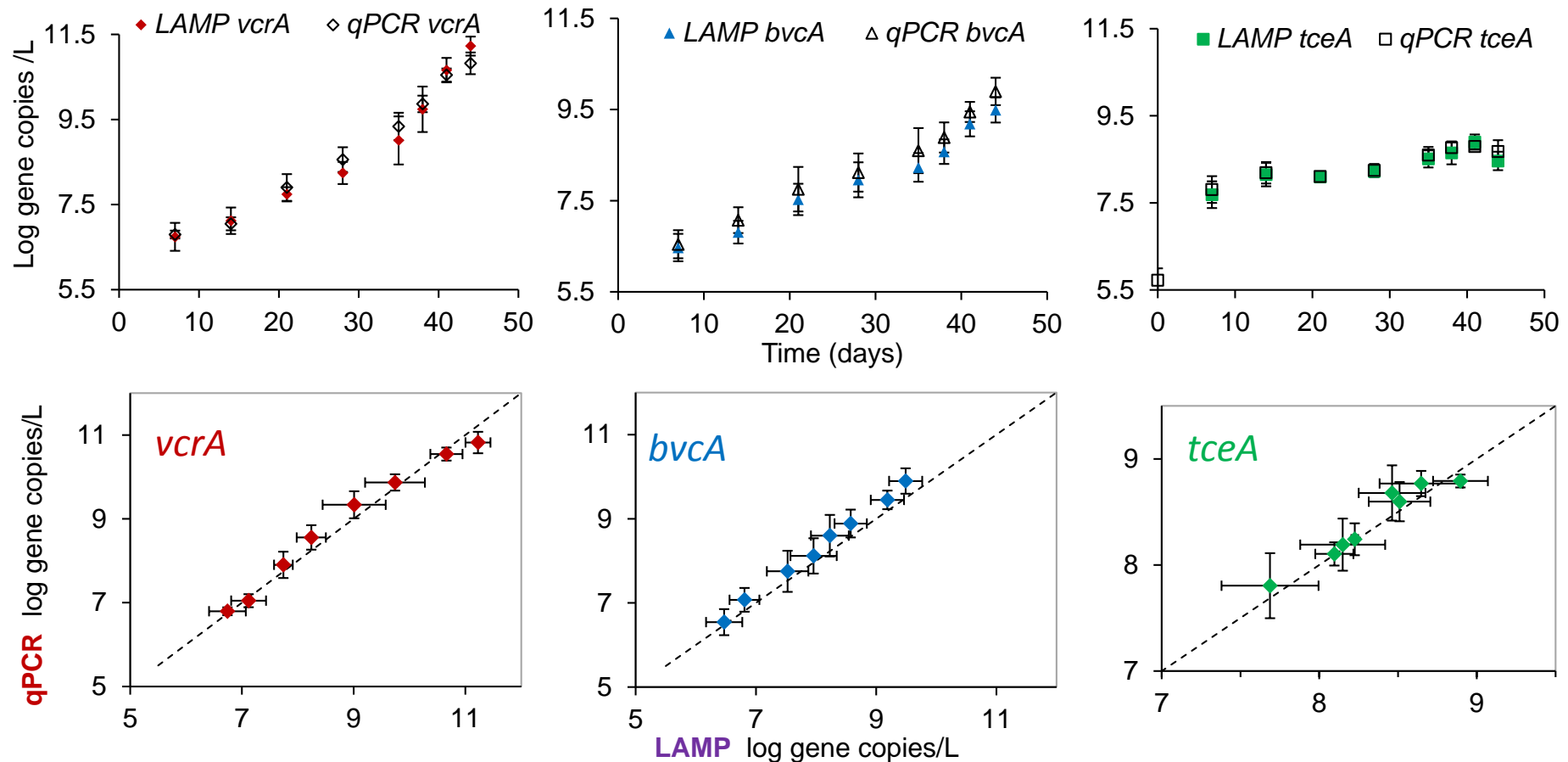
Q3. How viable are **visual based LAMP assays** on **cell templates** (no DNA extraction) for detecting RDase genes?





LAMP with Rdase genes

Comparing LAMP to qPCR with KB-1 DNA templates



- LAMP primers were used to track the growth of *Dehalococcoides* spp. over one growth cycle of KB-1
- Quantification with LAMP was comparable to qPCR
- Similar results were observed with SDC-9



LAMP with Rdase genes

Comparing LAMP to qPCR with groundwater templates

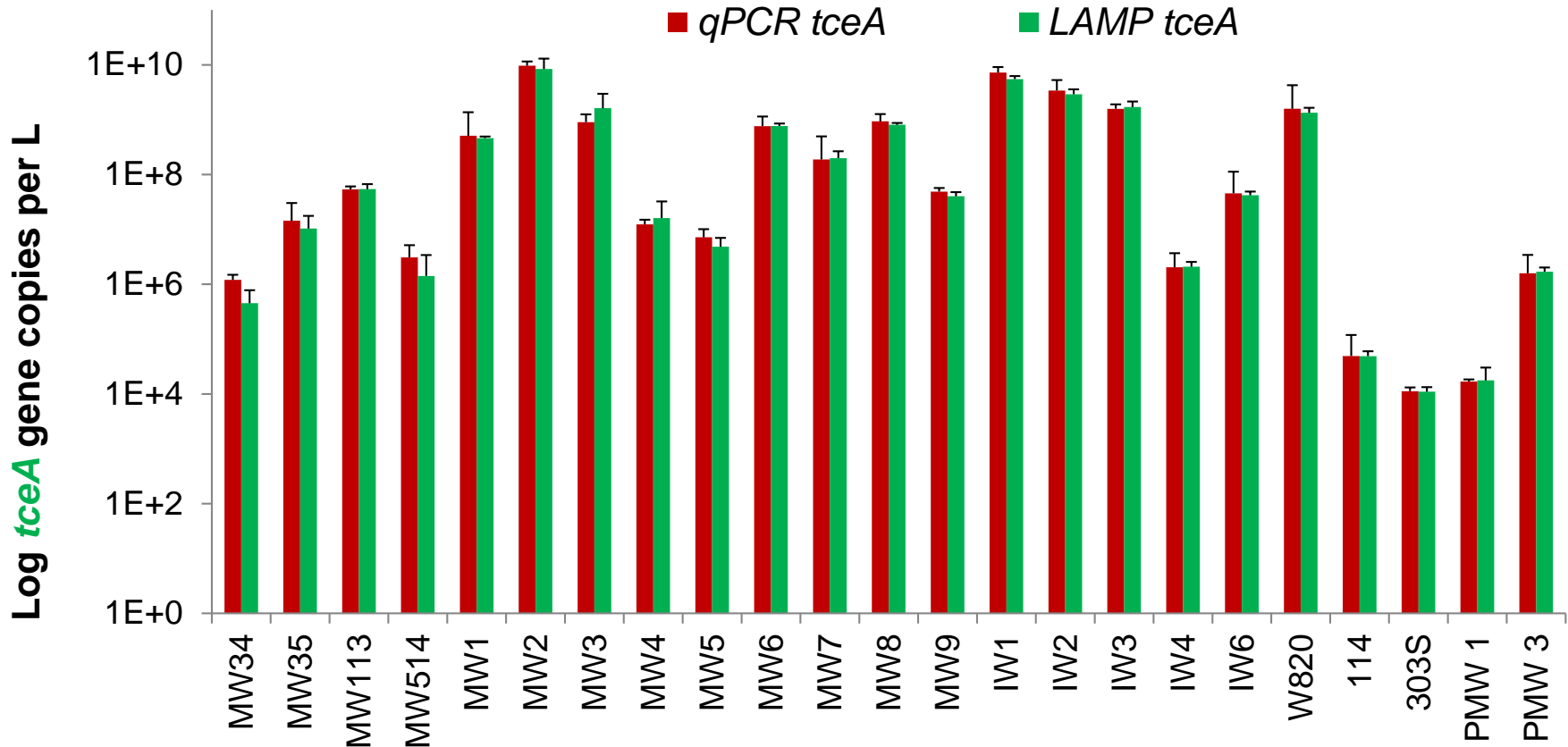


- 27 groundwater samples from 7 sites were shipped to Michigan State University for analysis
- DNA extracts from groundwater samples were used as templates for comparing LAMP to qPCR



LAMP with RDase genes

Comparing LAMP to qPCR with groundwater templates (*tceA*)

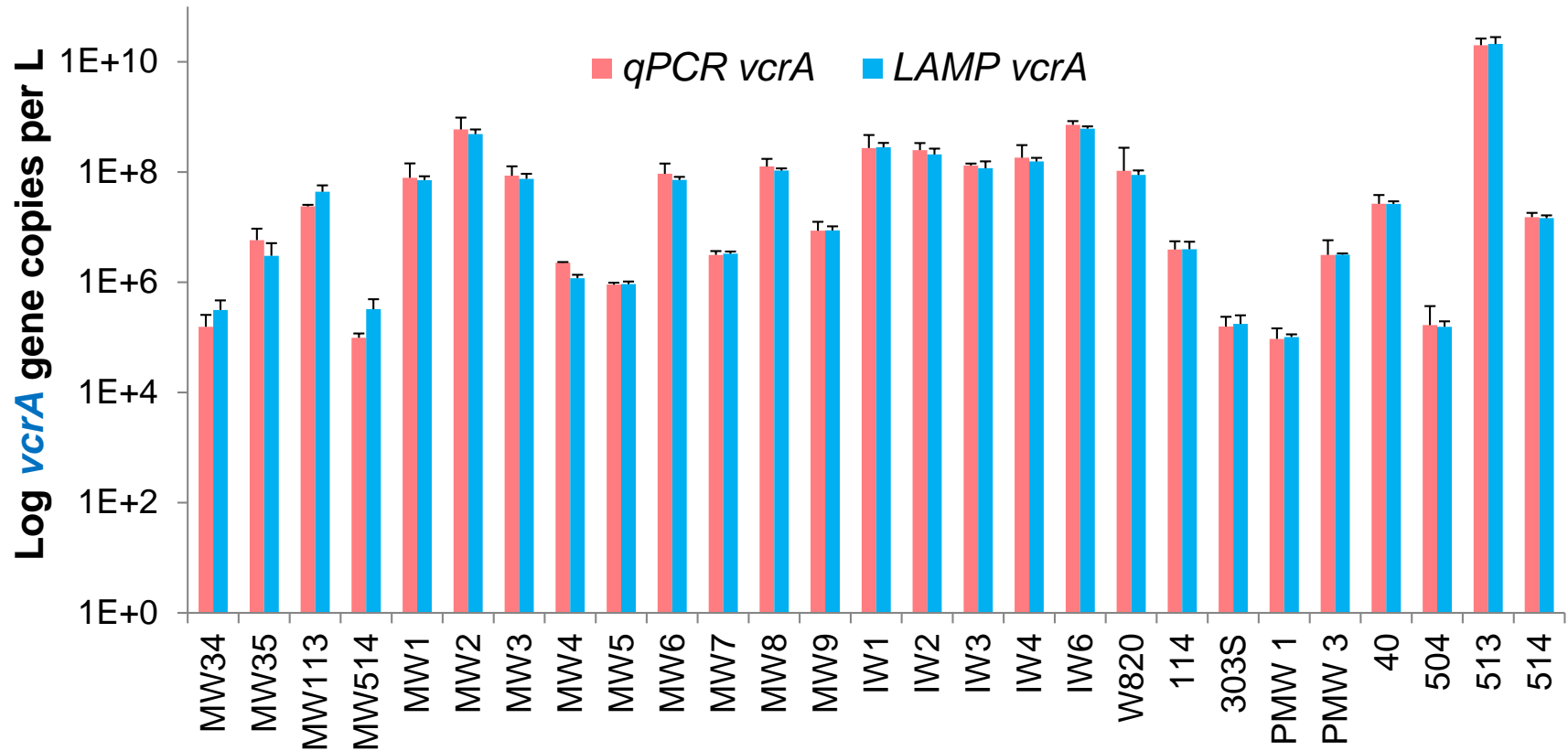


- qPCR & LAMP produced similar results for *tceA*
 - ✓ Similar results for different groundwater samples
 - ✓ Similar results over a range of concentrations
 - ✓ Replication was good between triplicate samples



LAMP with RDase genes

Comparing LAMP to qPCR with groundwater templates (vcrA)



- qPCR & LAMP produced similar results for *vcrA*
 - ✓ Similar results for different groundwater samples
 - ✓ Similar results over a range of concentrations
 - ✓ Replication was good between triplicate samples

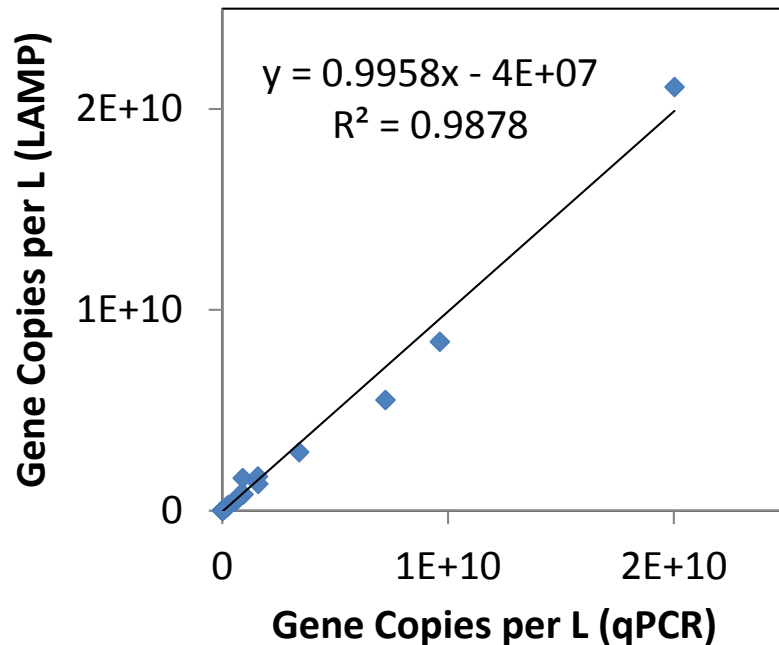


LAMP with RDase genes

Summary - DNA from Groundwater qPCR compared to LAMP

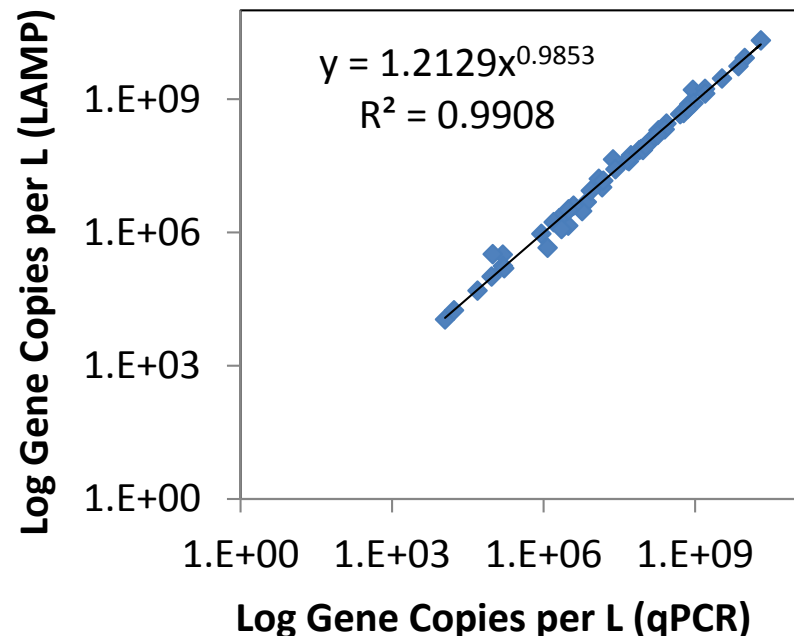
LAMP vs. qPCR (*tceA* & *vcrA*)

Arithmetic Scale



LAMP vs. qPCR (*tceA* & *vcrA*)

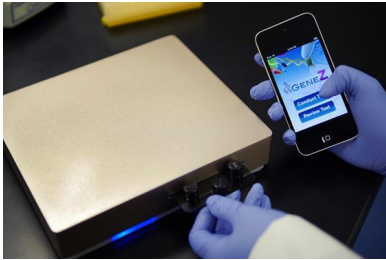
Log Scale



- qPCR & LAMP produced similar results for DNA extracted from cultures and from SDC-9 bioaugmented groundwater
- Quantification with LAMP was comparable to qPCR over a wide range of concentrations for *tceA* and *vcrA* genes when DNA was used as templates

Three Study Questions

Q1. Is **LAMP** comparable to qPCR for DNA extracted from cultures or groundwater?



Q2. Is **LAMP with Gene-Z** comparable to qPCR with a real time thermal cycler?

Q3. How viable are **visual based LAMP assays** on **cell templates** (no DNA extraction) for detecting RDase genes?

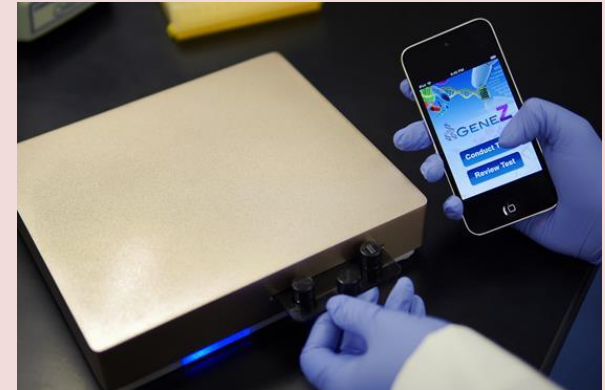




Alternate Platforms for Detection

Gene-Z Analyzer

- Gene-Z is an inexpensive handheld device developed by Dr. Hashsham's group for the parallel detection of pathogens
- It offers quantitative isothermal amplification based on LAMP and is operated using an iPod Touch
- Multiple LAMP assays can be performed on a microfluidic chip (made in house)
- The chip is loaded using a one step process for dispensing the sample
- Analysis of the sample usually takes less than two hours

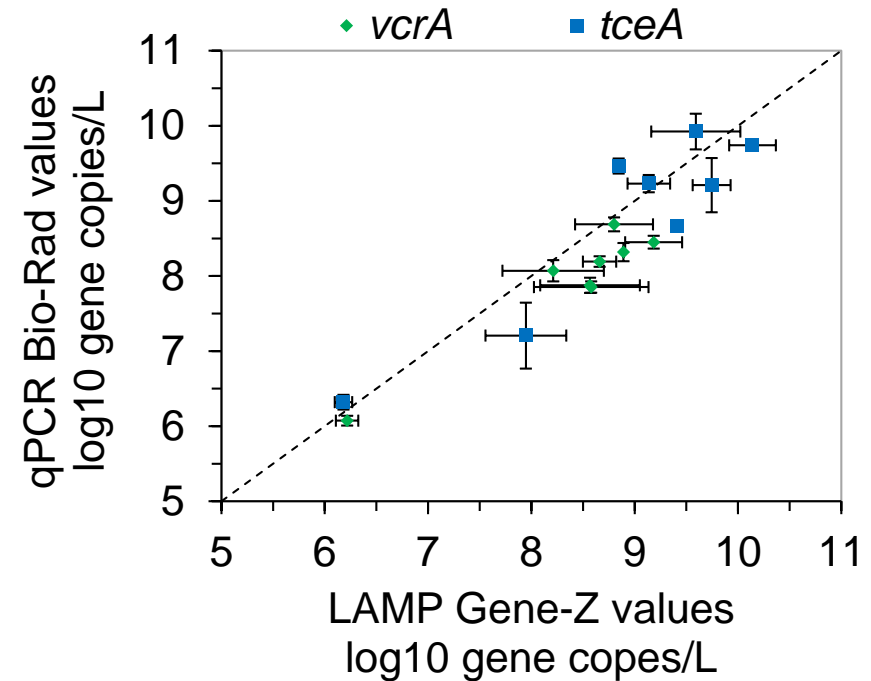




Alternate Platforms for Detection

Quantification in SDC-9 bioaugmented groundwater with LAMP

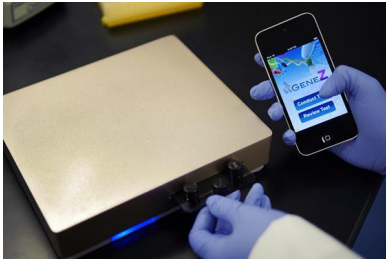
- DNA extracts from groundwater samples were used as templates to compare qPCR based quantification to quantification on the Gene-Z
- Quantification with LAMP Gene-Z was comparable to that of qPCR over a wide concentration range for both *tceA* and *vcrA* genes



Kanitkar, Y. H.; Stedtfeld, R. D.; Steffan, R. J.; Hashsham, S. A.; Cupples, A. M., Development of loop mediated isothermal amplification (LAMP) for rapid detection and quantification of *Dehalococcoides* spp. biomarker genes in commercial reductive dechlorinating cultures KB-1 and SDC-9. *Applied and Environmental Microbiology* **2016**, 82:1799-1806

Study Questions

Is LAMP comparable to qPCR for DNA extracted from cultures or groundwater?



Is LAMP with Gene-Z comparable to qPCR with a real time thermal cycler?

How viable are **visual based LAMP assays** on **cell templates** (no DNA extraction) for detecting RDase genes?





LAMP with RDase genes

Direct Cell Amplification on Real Time Thermal Cyclers

- Direct cell amplification is the **addition of cells without DNA extraction** to the LAMP reaction as a template for amplification
- It is very similar to **colony PCR** performed during M13 screen test
- Direct cell templates are often used with LAMP for **rapid point of care diagnostics** in human and veterinary medicine
- Direct cell and **centrifuged cell templates** were prepared from SDC-9 bio augmented groundwater and used to test LAMP

Creation of Direct and Centrifuged Cell Templates from Groundwater

Groundwater pre-filtered (5 μm), then passed through Sterivex filter



Add elution buffer (1000 μL), vortex to remove cells



Centrifuge, then resuspend the pellet



Direct Cell Templates

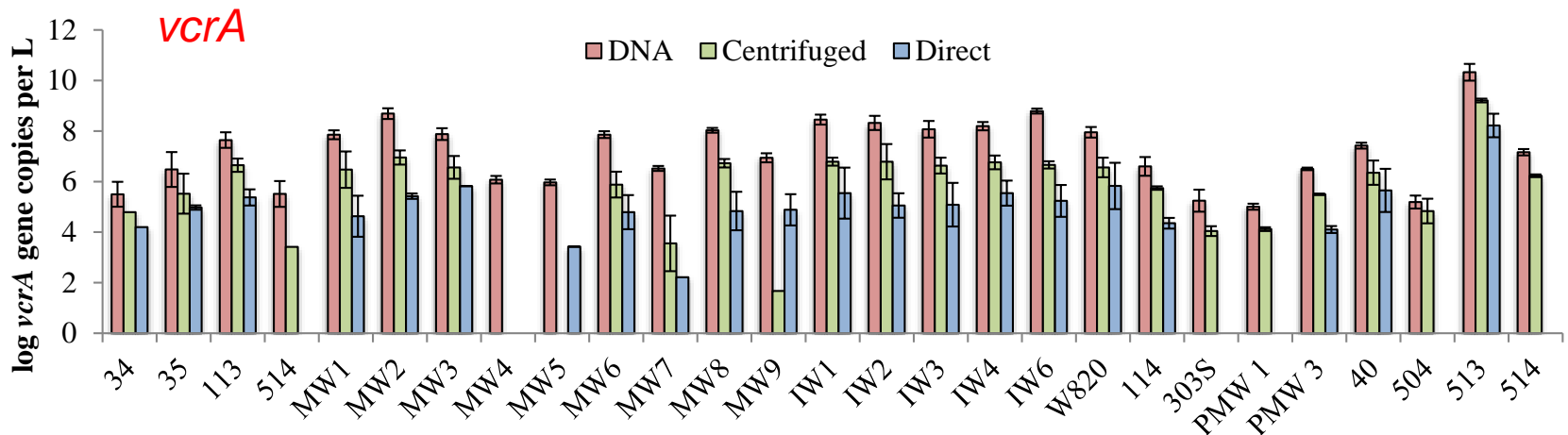
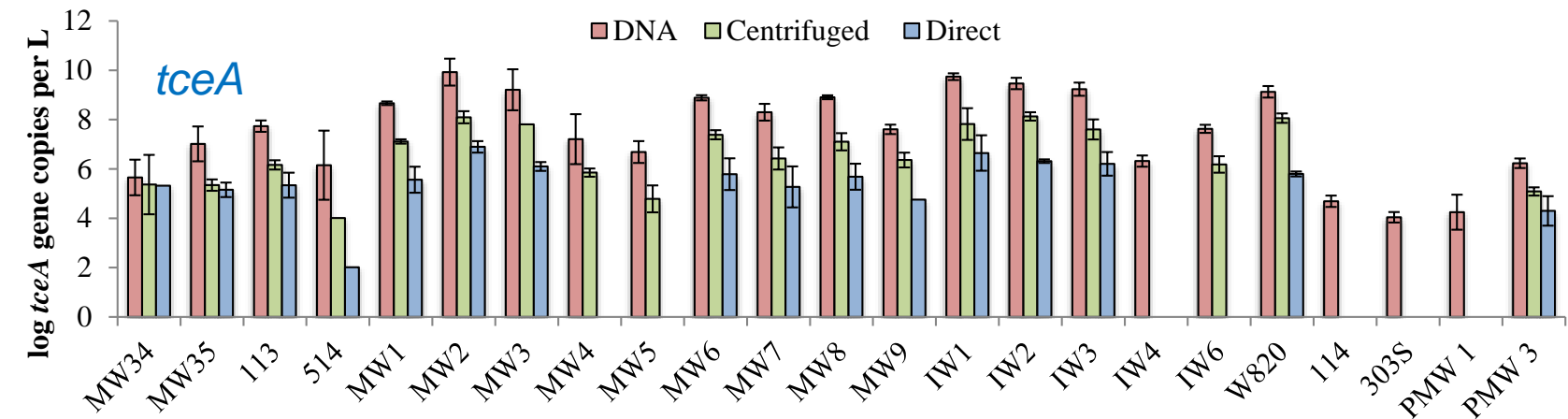
Centrifuged Cell Templates

Add 5 μL of sample to LAMP master mix



LAMP with RDase genes

Direct Cell Amplification on Real Time Thermal Cyclers



- Quantification with **DNA templates > Centrifuged cell templates > Direct cell templates**
- This trend was observed to be similar with both *tceA* and *vcrA* genes



LAMP with RDase genes

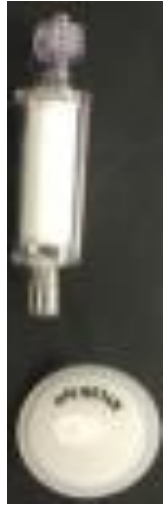
Summary - Amplification with Direct Cell Templates

- When biomass concentrations are increased using centrifugation, the observed gene copy/L concentrations also increase
 - This suggests that observed gene copy/L concentrations are a function of eluted biomass concentration
 - Quantification with centrifuged cell templates was more faithful to quantification with DNA templates
-
- Log gene copies/L (no DNA extraction)
- Log gene copies/L (with DNA extraction)
- Centrifuged
 Δ Direct
- $y = 1.7812x^{0.7953}$
 $R^2 = 0.918$
- $y = 0.125x^{0.771}$
 $R^2 = 0.687$
- Centrifuged cell templates had about half the gene copy concentration of DNA templates
 - Direct cell templates had $\sim 1/10$ gene copy concentration of DNA templates



Alternate Platforms for Detection

SYBR Green LAMP assay



Pre-filter (5 μm)
100 mL, then use
Sterivex filter



Elution buffer
(1000 μL), then
vortex & remove
cell



Centrifuge, then
resuspend pellet



Add 5 μL of sample
to LAMP master mix,
and then move to
water bath (1 hr,
63°C)



Add SYBR
Green



Orange: Control

Three Green: >1.8 X
10⁵ *vcrA* per L



Kanitkar Y.H., Stedtfeld R.D., Hatzinger P., Hashsham S.A., Cupples A.M. (In press) . Development and application of a rapid, user-friendly and inexpensive method to detect *Dehalococcoides* spp. reductive dehalogenase genes from groundwater. Applied Microbiology and Biotechnology



Alternate Platforms for Detection

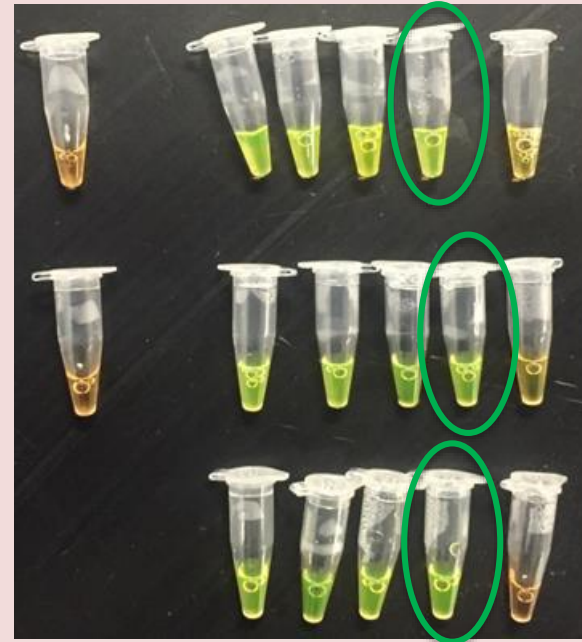
Detection Limits of SYBR Green LAMP Assay

- Detection limits of the SYBR Green LAMP assay when plasmid DNA is used as a template were evaluated
- A five fold 10X dilution series of *tceA*, *vcrA*, and *bvcA* genes were used as templates
- SYBR Green LAMP assay was able to detect **~100 *vcrA* gene copies per reaction** when plasmid DNA was used as a template for amplification
- Similar results were observed for *tceA* and *bvcA* genes

Detection limits with plasmid DNA (*vcrA* gene)

Controls Triplicates of a dilution series

10^5 10^4 10^3 10^2 10



Dilution series 1

Dilution series 2

Dilution series 3

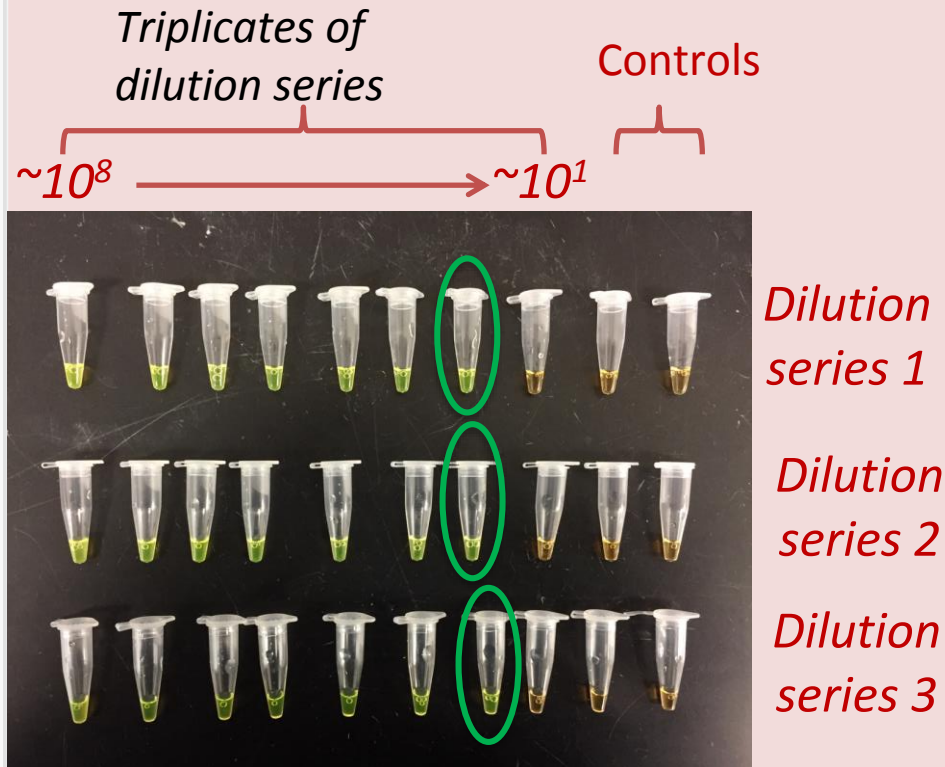


Alternate Platforms for Detection

Detection Limits with Direct Amplification

- Detection limits of the SYBR Green LAMP assay were evaluated with centrifuged cells prepared from diluted SDC-9 culture
- An eight fold 10X dilution series was containing $\sim 8.4 \times 10^7$ gene copies to ~ 8 gene copies were used as templates
- SYBR Green LAMP assay was able to detect **~ 84 *vcrA* gene copies per reaction** when centrifuged cells were used as a templates for amplification
- Similar results were observed for the *tceA* gene

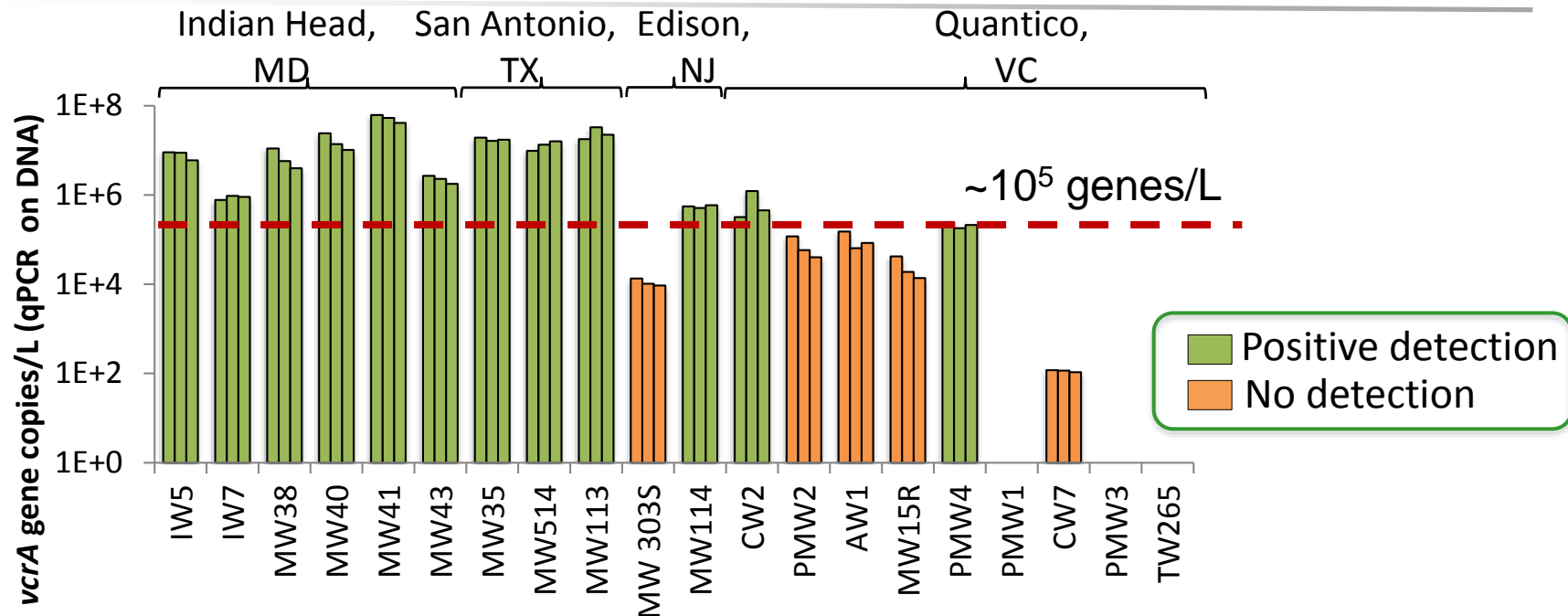
Detection limits with centrifuged cell templates (*vcrA* gene)





Alternate Platforms for Detection

Visual Detection of Centrifuged Cell Templates (Groundwater)



- SYBR Green LAMP assay were evaluated with centrifuged cells prepared from groundwater samples from 4 different sites.
- LAMP assay detected *vcrA* gene above $\sim 10^5$ genes/L
- $>10^6$ gene copies/L required for efficient dechlorination*

If all three replicates remain orange, bioaugmentation or biostimulation may be necessary

*Lebrón, C. A., E. Petrovskis, F. Löffler & K. Henn, Jan 2011, Guidance Protocol, ER-0518

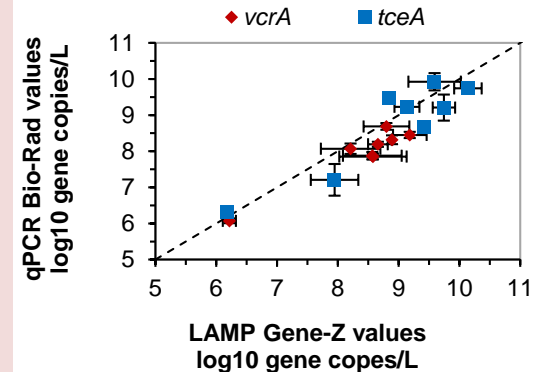
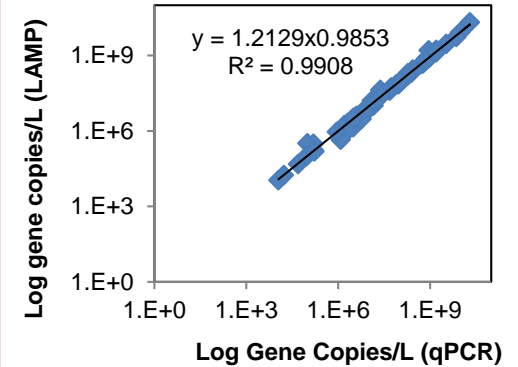
Kanitkar, Y. H., Stedtfeld, R. D., Hatzinger, P. B., Hashsham, S. A., and A. M. Cupples. *In Press*. Development and application of a rapid, user-friendly and inexpensive method to detect *Dehalococcoides* sp. reductive dehalogenase genes from groundwater. *Applied Microbiology & Biotechnology*.



Conclusions

Lessons Learned and Current Research

- LAMP and qPCR produced similar results when DNA templates were used.
- This suggests that LAMP can be used to track the growth of *Dehalococcoides* spp. in cultures and groundwater
- LAMP in Gene Z produced similar results as qPCR in a thermal cycler
- This suggests that LAMP can be performed on alternative platforms and may not necessarily require real time thermal cyclers
- Visual based LAMP assay has the potential for field deployment
- Current research focuses on making the LAMP based assay quantitative (MPN)





Acknowledgements

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- **Phil Dennis from SiREM** for supplying KB-1 culture
- **Frank Löffler from University of Tennessee at Knoxville** for providing the *tceA* plasmid standard