Kinetic Analysis Implicates Nitrous Oxide as a Potent Inhibitor of the Bacterial Reductive Dehalogenation Process

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Background/Objectives. Intense usage of nitrogen (N) fertilizer has resulted in increased nitrate concentrations in groundwater. Under anoxic conditions, nitrate is reduced via microbial denitrification to nitrous oxide (N₂O) and dinitrogen (N₂). As a consequence, groundwater N₂O concentrations exceed the equilibrium concentration with atmospheric N₂O (~7 nM at 20 °C) by up to 4 orders of magnitude. N₂O reacts with certain metalloorganic prosthetic groups such as cobalt- (Co-) containing corrinoids (e.g., vitamin B₁₂). The chemical reaction of N₂O with Co(I) can cause damage to corrinoid-dependent enzyme systems and thus impact an organism's metabolism. Organohalide-respiring bacteria require corrinoid for assembling functional reductive dehalogenases, which are the key enzyme system catalyzing reductive dechlorinates tetrachloroethene (PCE) to *cis*-1,2-dichloroethene (*c*DCE), and *Dehalococcoides mccartyi* strain BAV1 dechlorinates *c*DCE via vinyl chloride (VC) to environmentally benign ethene. As global fertilizer usage increases, the impact of elevated N2O concentrations on relevant microbial processes in (contaminated) aquifers must be understood.

Approach/Activities. The effect of N₂O ranging from 5 to 100 μ M on the corrinoid-dependent reductive dechlorination activities in *Geobacter lovleyi* strain SZ and *Dehalococcoides mccartyi* strain BAV1 were examined in whole cell suspension assays. A Michaelis-Menten single substrate model was used to illustrate the inhibitory effects of N₂O on the reductive dechlorination process.

Results/Lessons Learned. Low concentrations (9.5 µM) of N₂O impacted PCE and cDCE reductive dechlorination rates and extents in strain SZ and in strain BVA1 cultures, respectively, suggesting that N₂O is a potent inhibitor of corrinoid-dependent reductive dechlorination. The most pronounced inhibition was observed for the VC reductive dechlorination step in strain BAV1 cultures, and only negligible amounts of VC were dechlorinated at N₂O concentrations exceeding 19 µM. Following removal of 95 µM N₂O from inhibited cultures, the reductive dechlorination activity was fully restored in strain SZ cultures. In contrast, recovery of VC dechlorination activity was not observed in strain BAV1 cultures even after extended incubation periods when the initial N₂O concentration exceeded 29 µM. The kinetic parameters for both isolates in the presence of 0-60 μ M N₂O were determined using the Michaelis-Menten model. In strain SZ assays, the whole-cell half-saturation constant (K_s) of 25.1±2.9 μ M for PCE was not affected by 60 μ M N₂O, but ~26 μ M N₂O reduced the maximum PCE dechlorination rate V_{max} of 76.3 \pm 2.6 nmol CI- released/min/mg protein by 50%. A measured inhibitory constant (K) of 26.0 \pm 4.9 µM indicated that PCE dechlorination rate (i.e., Vmax) is impacted at environmentally relevant N₂O concentrations. Similar results were observed for *c*DCE dechlorination in strain BAV1 assays. In the absence of N₂O, a Ks value of 23.1±4.1 µM and a Vmax value of 121.3±6.2 nmol CI- released/min/mg protein were determined. When exposed to 17.7-24.7 µM N₂O, the Vmax of cDCE dechlorination decreased ~50%, and a K_i value of 21.2±3.5 μ M was determined. These kinetic measurements highlight that N₂O inhibition of reductive dechlorination must be expected at environmentally relevant concentrations, particularly in aguifers receiving agricultural nitrate runoff. Considering the commonality of elevated nitrate concentrations in many aquifers, the findings provide an explanation for stalled or incomplete reductive dechlorination at sites impacted with chlorinated pollutants. Thus, groundwater monitoring regimes should include nitrate and N₂O measurements, so that potential inhibition and VC stalls can be explained and predicted.