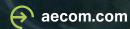


Natural Occurrence of Feammox Conditions and Anammox Microbiota within a PFAS Plume at the Ground Water-to-Surface Water Interface

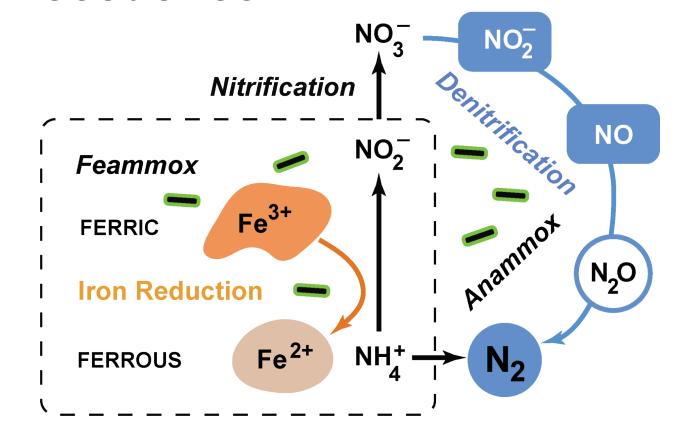
Barry J. Harding, Director of Nature-Based Solutions
AECOM Remediation West





Feammox Conditions / Precedence

- Feammox = Oxidation of ammonia under iron reducing conditions
- Feammox also refers to bacteria associations
- Anammox = Anaerobic ammonium oxidation and associated bacteria
- Acidimicrobium sp. Strain A6, encoded for novel dehalogenases and studied for defluorination capabilities (Jaffé and others)



Interesting Bacteria

Feammox reaction modified from graphic derived from Jaffé, Huang, Park and Koel (Princeton U), April 15, 2022.





Purpose of Study

- Field studies documenting Feammox conditions are lacking
 - Studies to-date were in flooded rice paddies, manure composts, and bacteria harvesting in the New Jersey Pine Barrens
- Most Feammox studies are in-vitro (bench-scale reactor vessels and controlled experimentation)
- This study focused on documenting Feammox conditions in situ within a PFAS-containing groundwater-to-surface water discharge

Goal

 Document if Feammox conditions and related bacteria are present within a PFAS-laden groundwater seep to surface water and identified study area





Field Study Design

- Pore-Water Sampling and Analysis
 - Three push-point samples
 - Low flow minimal drawdown and use of peristaltic pump
 - pH, ORP, Sp.Cond, DO
 - Feammox indicators: Ammonia, Iron, Ferrous Iron, Nitrate-N, Nitrate-N
 - PFAS-28; Two sampling events
- Sediment Sampling and Analysis
 - Three co-located hand-auger borings
 - PFAS-28
 - NGS, 16s rRNA Amplicon, preserved with DNA/RNA Shield® (Zymo Research)





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Study Area

- Within an identified dissolved PFAS discharge adjacent to a Great Lakes tributary
- Stream was gaining during sampling activities in September 2022 and January 2023
- Iron-rich sediments including hematite, magnetite, goethite and various iron-hydroxides
- Study depth limited to < 3-feet below grade and consistent with known PFAS occurrences adjacent to tributary

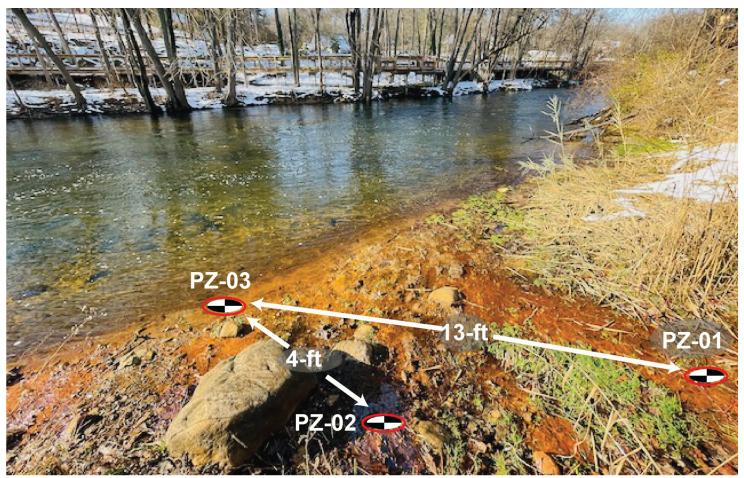


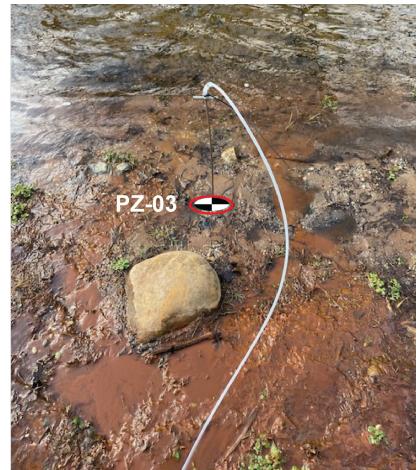






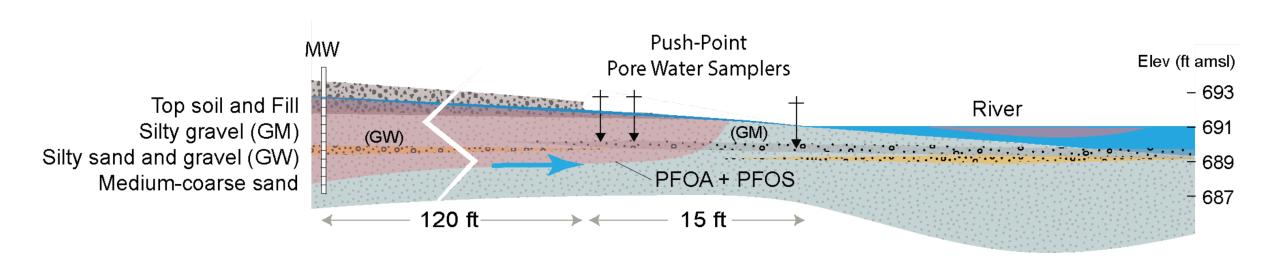
Pore-Water Sampling Locations







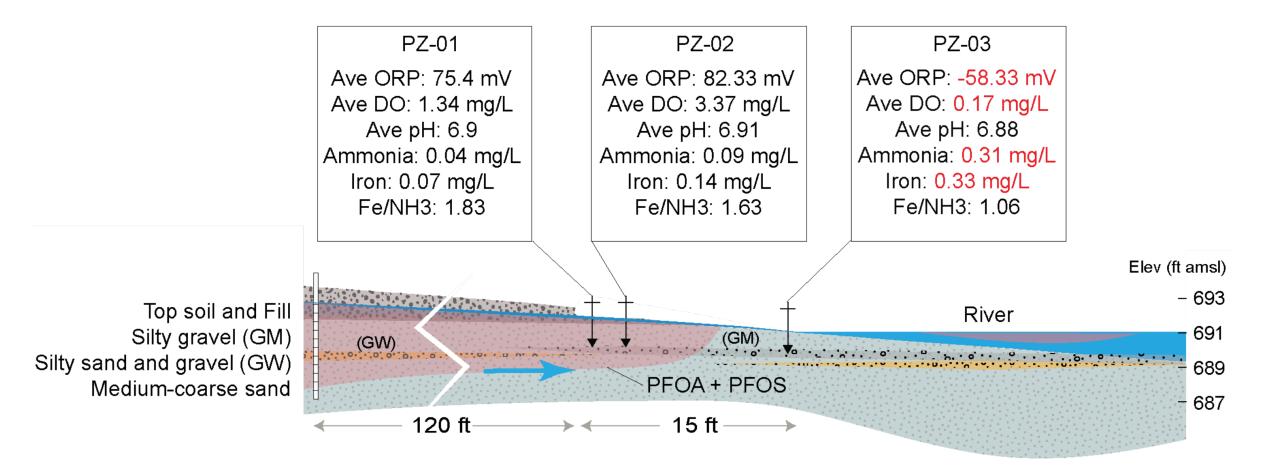
Study Area Cross-Sectional View







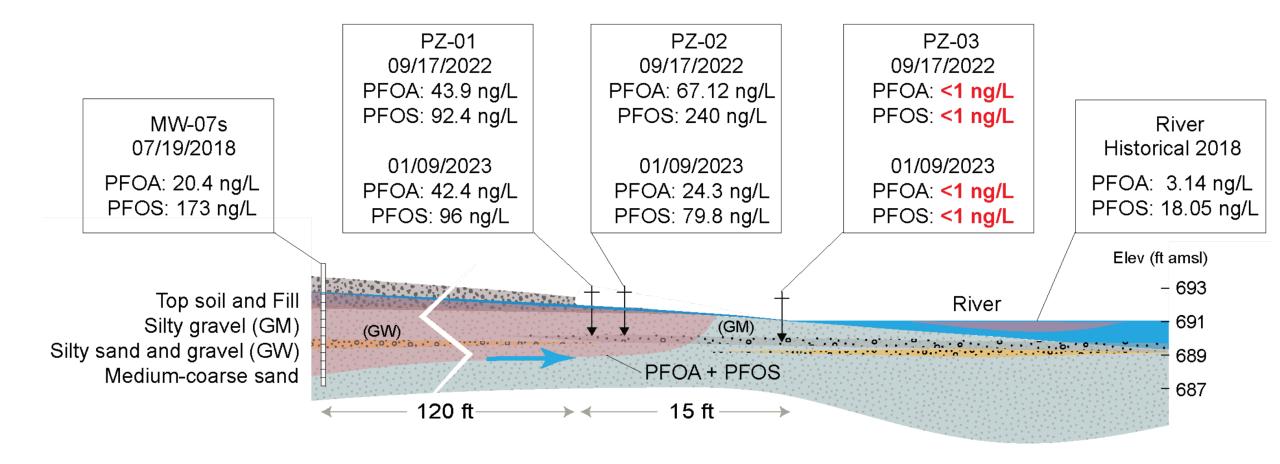
Water Quality & Feammox Indicators







PFAS Sampling Results





16S Amplicon Sequencing

- Nine sediment samples collected from each pore-water location with three discrete sampling depths (0.5, 1.0 and 1.5-foot intervals)
- Samples preserved in field with DNA/RNA® Shield
- Microbiomics Reporting, Zymo Research, Irvine California
 - ZymoBIOMICS® 96 MagBead DNA Kit
 - Quick-16S™ Primer Set V3-V4
 - Sequencing: Illumina® MiSeq™ v3 Reagent
 - Bioinformatics Analysis: Dada2 Pipeline





16S rRNA Amplicon Sequencing - ORDER LEVEL

PZ-01 PZ-03

	Taxon - Order	Gene %		Taxon - Order	Gene %		Taxon - Order	Gene %
1	Rhizobiales	11.54%	1	Rhizobiales	10.52%	1	Anaerolineales	7.16%
2	Burkholderiales	6.07%	2	Nitrosomonadales	5.54%	2	Rhizobiales	6.99%
3	Acidimicrobiales	5.33%	3	Class Acidobacteria; Order NA	5.48%	3	Chloroflexi; Class and Order NA	6.07%
4	Chloroflexi; Class and Order NA	5.30%	4	Acidimicrobiales	5.33%	4	Acidobacteria; Order NA	4.89%
5	Rhodobacterales	3.96%	5	Chloroflexi; Class and Order NA	4.49%	5	Deltaproteobacteria; Order NA	4.02%
6	Xanthomonadales	3.87%	6	Nitrospirales	4.29%	6	Burkholderiales	3.68%
7	Acidobacteria; Order NA	3.55%	7	Burkholderiales	4.06%	7	Nitrospirales	3.54%
8	Nitrospirales	3.15%	8	Xanthomonadales	3.66%	8	Acidimicrobiales	3.53%
9	Anaerolineales	3.00%	9	Anaerolineales	3.33%	9	Gaiellales	2.65%
10	Nitrosomonadales	2.56%	10	Deltaproteobacteria; Order NA	2.59%	10	Xanthomonadales	2.49%
		48.35%			49.28%			45.03%
	Other Taxon	51.65%		Other Taxon	50.72%		Other Taxon	54.97%

Rhizobiales:

Broad group of root-zone dwelling bacteria that fix atmospheric nitrogen and are involved in nitrogen cycling **Anaerolineales**:

Filamentous, fermentative, strictly anaerobic chemo-organo-heterotrophs





16S rRNA Amplicon Sequencing – GENUS LEVEL

PZ-01

	Taxon - Genus	Gene %
1	Nitrospira	1.86%
2	Acidiferrobacter	1.26%
3	Illumatobacter	1.10%
4	Gaiella	0.99%
5	Rhodobacter	0.96%
6	Hyphomicrobium	0.96%
7	Hydrogenophaga-Variovorax	0.88%
8	Tabrizicola	0.87%
9	Nocardioides	0.83%
10	Pedomicrobium	0.82%
11	Leptothrix	0.72%
12	Methylomirabilis	0.68%
13	Amaricoccus	0.68%
14	Denitratisoma	0.65%
15	Luteolibacter	0.54%
16	Alysiosphaera	0.52%
17	Haloferula	0.50%
18	Nordella	0.50%
19	Devosia	0.46%
20	Sphingomonas	0.43%

PZ-02

	Taxon - Genus	Gene %
1	Nitrospira	2.45%
2	Hyphomicrobium	1.51%
3	Illumatobacter	0.94%
4	Leptothrix	0.93%
5	Methylomirabilis	0.86%
6	Variibacter	0.84%
7	Nitrotoga	0.68%
8	Ornithinibacter	0.66%
9	Nitrosomonas	0.66%
10	Denitratisoma	0.65%
11	Rhodoferax	0.65%
12	Gaiella	0.60%
13	Acidiferrobacter	0.59%
14	Methylobacter	0.57%
15	Nordella	0.56%
16	Nocardioides	0.52%
17	Bryobacter	0.51%
18	Nakamurella	0.49%
19	Amaricoccus	0.48%
20	Pedomicrobium	0.47%

PZ-03

	Taxon - Genus	Gene %
1	Hyphomicrobium	1.93%
2	Gaiella	1.29%
3	Leptothrix	1.14%
4	Nitrospira	0.65%
5	Brocadia	0.50%
6	Bryobacter	0.46%
7	Nocardioides	0.46%
8	Pedomicrobium	0.45%
9	Clostridium	0.44%
10	Acidiferrobacter	0.43%
11	Desulfatiglans	0.43%
12	Methylomirabilis	0.38%
13	Desulfobacca	0.37%
14	Ignavibacterium	0.37%
15	Thiobacillus	0.36%
16	Rhodobacter	0.36%
17	Denitratisoma	0.35%
18	Haloferula	0.34%
19	Variibacter	0.33%
20	Luteolibacter	0.32%

ANAMMOX BACTERIA

DENITRIFYING BACTERIA

"IRON" BACTERIA

DCM DEHALOGENASE





Feammox-Like? **Microbes** Reducing Conditions Ferrous Fe **Feammox** 6:1 Iron Fe-NH3 **Minerals** N Cycling



Yellow = Condition Present at Study Area



Conclusions

- Feammox-like conditions were verified in the field through a weight of evidence including presence of iron mineralization, ammonia, reducing conditions (at the groundwater-to-surface water interface at PZ-03) and specific bacteria, including *Nitrospira*, *Brocadia*, and *Denitratisoma*.
- Optimal iron to ammonia ratio (6:1) for Feammox were not present in the study area
- One identified bacterium Hyphomicrobium sp. is included within a genus documented as being encoded with dehalogenase genes.
- PFAS were conspicuously absent during all sampling events at location PZ-03, which yielded the most representative Feammox conditions (redox and microbial footprint).





Next Steps

- Further temporal monitoring is warranted with additional sampling and analysis for Feammox indicators and PFAS
- Additional use of Molecular Biological Tools (MBTs) will focus on identifying functional genes associated with the promising bacteria
- Focused sampling and bench-scale microcosm culturing of study area sediments







Team Acknowledgements

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- Wes Stambaugh, Field Data Collection
- Rosa Gwinn, Oversight and Chief Morale Officer
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